Case Report: Disseminated Autochthonous Dermal Leishmaniasis Caused by Leishmania siamensis (PCM2 Trang) in a Patient from Central Thailand Infected with Human Immunodeficiency Virus

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INTRODUCTION

Leishmaniasis is a rare tropical disease found in tropical and subtropical countries. Thailand was once considered to be a leishmaniasis-free country. However, during 1960–1986, a few cases of leishmaniasis were described among Thai people who had been to the endemic areas. Since 1996, sporadic indigenous cases of leishmaniasis were reported in 1996, 2005, and 2007. In 2008, a new causative species of autochthonous visceral leishmaniasis Leishmania siamensis was described for the first time by Sukmee and others. From then on, more than 10 cases of autochthonous visceral leishmaniasis caused by L. siamensis have been occasionally reported. Until recently, it has been evident that most of the so-called L. siamensis are Leishmania martiniquensis.

To date, there has been only one published case of PCM2 in southern Thailand. Here we report, for the first time, L. siamensis (PCM2 Trang isolate) from a patient living in central Thailand. Furthermore, by large subunit of RNA polymerase II gene and ribosomal RNA internal transcribed spacer 1 (ITS-1) sequencing, we demonstrate that this Leishmania appears to be identical or very similar to an organism previously reported from Trang Province of southern Thailand, namely PCM2 isolate.

CASE REPORT

In July 2015, a 42-year-old human immunodeficiency virus (HIV)-infected Thai woman with a CD4 count of 89/mm³ presented with 2 weeks of low grade fever and multiple painless erythematous to hyperpigmented plaques and nodules on the skin of her face and lower extremities. These lesions gradually increased in size and number during observation over 3 months.

She was a housewife who lived in the Kanchanaburi Province of central Thailand and had never traveled outside Thailand and other parts of Thailand apart from central area.

Ten months ago, she was diagnosed with HIV infection presenting with a CD4 count of 9/mm³ and has had opportunistic infections of Pneumocystis jiroveci pneumonia and pulmonary tuberculosis. Her medications include stavudine, lamivudine, ritonavir-boosted lopinavir, efavirenz, cotrimoxazole, and fluconazole. Physical examination showed sign of wasting and body temperature of 37.6°C. The patient had moderate pallor with no jaundice. The examination of abdomen showed mild splenomegaly. Her CD4 T-cell count was 89 cells/mm³. Hypoalbuminemia of 7.9 g/dL, a white blood cell count of 5,400 cells/mm³ (neutrophils 71%, lymphocytes 16%, eosinophils 5%, basophils 0%, and monocytes 8%), and a platelet count of 422,000 cells/mm³. Liver function tests gave the following results: an aspartate aminotransferase level of 72 U/L (7–48 U/L), and an alkaline phosphatase level of 314 U/L (45–115 U/L). Her CD4 T-cell count was 89 cells/mm³. Hypoalbuminemia (2.4 g/dL) and hypergammaglobulinemia (6.6 g/dL) were also noted. Other laboratory results were within normal limits. A cutaneous biopsy was performed and histopathology demonstrated normal epidermis. Many nonencapsulated intracytoplasmic yeast-like organisms were noted in the dermis with multiple patchy infiltration of mixed inflammatory cells, particularly, lymphocytes, histiocytes, and neutrophils. Binary fission was not seen (Figure 2). All the special stains such as periodic acid–Schiff stain, Giemsa stain, and Gomori methenamine silver stain showed a positive result. Ultrasonography of abdomen revealed mild splenomegaly. Bone marrow study was normal.

After culture on Sabouraud dextrose agar for 2 weeks showed no fungal growth, molecular characterization was performed from tissue biopsy using polymerase chain reaction (PCR) amplification and sequencing and comparing the

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database from GenBank. Pan-fungal PCR for ITS-1 and ITS-2 appeared negative but positive for D1/D2. Subsequently, DNA sequencing was done for D1/D2. The sequences revealed 99% homology to *Leishmania amazonensis* (accession no. JX030045). Therefore, another PCR and DNA sequencing were performed using primer specific to *Leishmania* for its identification. Sequences from two independent loci were examined: the ITS-1 region of the 18S ribosomal RNA gene and the large subunit of RNAPolI. The large subunit of RNAPolII sequences of our parasite appeared to be 1,181/1,184 base pairs (99%) identical to *L. siamensis* previously reported (GenBank accession no. KM820664) while ITS-1 sequence showed 282/295 base pairs (96%) similarity (JX195640). GenBank accession nos. for the new sequences are KX347438 for ITS-1 and KX347439 for RNAPolII.

The sequences obtained from our case and PCM2 from Trang (JX195640) were the most similar while lower identity (95%) of ITS-1 sequence was observed between a pair of our *Leishmania* and Ghanaian *Leishmania* (EF524071). In addition, the lower identity (98%) of RNAPolII sequence between our strain and *Leishmania enriettii* (AF151727) was observed. Therefore, we concluded that patient was infected with *L. siamensis* (PCM2 Trang). Intravenous amphotericin B deoxycholate 0.6 mg/kg/day was administered daily for 4 weeks, and then she was referred back to Samutsakorn general hospital for further treatment with intravenous amphotericin (total dose 1,260 mg) followed by itraconazole 400 mg/day indefinitely due to the patient’s immunocompromised status. Her general condition was improved, and her skin lesions decreased in size at the time of referral.

**DISCUSSION**

To the best of our knowledge, this is the second report of PCM2 isolate. However, the name “*Leishmania siamensis*” remains confusing due to mistaking *L. martiniquensis* for *L. siamensis* in the past. In 2013, Leelayoova and others discovered that the so-called *L. siamensis* has indeed two separate lineages: the TR lineage and PG lineage. Further works by Pothirat and others clarified using sequence
analysis that a novel isolate from northern Thailand LSCM1 (accession no. JX899838) and all other strains except PCM2 Trang were L. martiniquensis (PG lineage), as previously stated from the Martinique island. This reflected that L. martiniquensis is the most common cause of autochthonous leishmaniasis in Thailand. Therefore, we can draw conclusion that autochthonous leishmaniasis in Thailand caused by two species of Leishmania: L. martiniquensis and L. siamensis. Although L. martiniquensis has worldwide distribution, our current strain appeared more restricted to Thailand. However, due to small number of PCM2 cases, further reports need to be collected for further clarification of various strains of Leishmania.

As Thailand is an endemic area for penicilliosis and other fungal HIV infections, the culture for leishmaniasis is not routinely performed in such cases. In terms of molecular diagnosis, we used primers designed from loci previously used for Leishmania identification. ITS-1 region is the most variable marker that can be used to analyze the genetic difference among Leishmania species.

The clinical manifestations of novel leishmaniasis in Thailand can be categorized into three clinical forms: visceral, diffused cutaneous, and diffused cutaneous combined with visceral. The infection has been mostly reported in immunocompromised patients, particularly those with HIV infection. Other than L. martiniquensis and L. siamensis, Leishmania infantum was also reported to be the cause of autochthonous leishmaniasis in Thailand. Our patient had low-grade, subacute and disseminated cutaneous lesions without other visceral involvement.

The variety of clinical presentations of cutaneous leishmaniasis does not depend only on the causative strains, but also on the nature of the host immune status, exclusively immune suppression caused by HIV infection. It is possible that PCM2 isolate is uniquely able to cause these cutaneous manifestations in patients with acquired immune deficiency syndrome. In immunocompetent hosts, a T-helper cell 1 (Th1) response provides protective effect against Leishmania, whereas Th2 cytokines causes susceptibility to infection and disease progression. HIV infection facilitates a Th2 immune response that promotes the progression of leishmaniasis.

Recently, cavernoculous species of phlebotomine sand flies from Kanchanaburi Province has been studied providing the diversity of sand flies potential to be leishmaniasis vector. However, limitation of this study was identification of sand flies vector. Epidemiological studies are necessary to determine life cycle of novel Leishmania with potential vectors and reservoir hosts for developing further disease control and prevention in Thailand.

In summary, this is the report of disseminated autochthonous cutaneous leishmaniasis in central Thailand, and the causative organism is identified as L. siamensis (PCM2 Trang). It is evident that autochthonous leishmaniasis in Thailand is caused by two species of Leishmania: L. martiniquensis and L. siamensis. Our report supports the existence of L. siamensis (PCM2 Trang). Although, Thailand is not regarded as endemic area for leishmaniasis, diffuse cutaneous infiltration with yeast-like organism in histopathology especially in immunocompromised patient should raise the suspicion of cutaneous leishmaniasis. Moreover, molecular diagnosis has been proved to be fast and effective method for diagnosis of leishmaniasis.

References


Note: Supplemental information appears at www.ajtmh.org.

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