Entomologic Inoculation Rates of *Anopheles arabiensis* in Southwestern Ethiopia

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**Abstract.** We collected anophelines every second week for one year from randomly selected houses in southwestern Ethiopia by using Centers for Disease Control (CDC) light traps, pyrethrum spray catches, and artificial pit shelter constructions to detect circumsporozoite proteins and estimate entomologic inoculation rates (EIRs). Of 3,678 *Anopheles arabiensis* tested for circumsporozoite proteins, 11 were positive for *Plasmodium falciparum* and three for *P. vivax*. The estimated annual *P. falciparum* EIR of *An. arabiensis* was 17.1 infectious bites per person per year (95% confidence interval = 7.03–34.6) based on CDC light traps and 0.1 infectious bites per person per year based on pyrethrum spray catches. The *P. falciparum* EIRs from CDC light traps varied from 0 infectious bites per person per year (in 60% of houses) to 73.2 infectious bites per person per year in the house nearest the breeding sites. Risk of exposure to infectious bites was higher in wet months than dry months, with a peak in April (9.6 infectious bites per person per month), the period of highest mosquito density.

**INTRODUCTION**

Although recent trends show a reduction in the prevalence of malaria in Ethiopia, it is still a challenge to the health of many communities. Long-lasting insecticidal treated nets (LLINs) and indoor residual spraying (IRS) are the two main malaria vector control tools being used to decrease mosquito density and longevity, and human–vector contact of *Anopheles arabiensis*, the species responsible for >95% of malaria transmission. The impact of LLINs and IRS on indoor populations of vectors is to reduce entomologic transmission indicators, the most common of these is the entomologic inoculation rate (EIR), which is the product of sporozoite rate and human biting rate over a defined time and space. Parity (longevity), sporozoite rates, and human blood index. The success of both interventions depends on the response of vectors, their behavior and interaction with humans, and alternative hosts. The EIR measures the intensity of malaria transmission in a particular area. Estimation of the EIR is important for quantifying the potential level of human exposure to infected mosquitoes and for assessing the impact of interventions on malaria transmission in a given area. Many studies have reported huge variations in malaria transmission intensity in Africa, not only between urban and rural settings but even within the same locality. It has been reported that the EIR varies between 0.01 and 1000 infectious bites per person per year (ibp/pyear) in malaria-endemic countries in Africa. Infections by *An. arabiensis* have been reported for highland villages with unstable malaria transmission in the south-central region of the country. Thus, there are substantial gaps in knowledge regarding entomologic transmission levels and the impact of the current anti-vector operations (IRS and LLINs).

Malaria is clearly a public health problem in Chano village in southwestern Ethiopia. In the absence of entomologic information on malaria transmission in this village and the surrounding areas, a detailed longitudinal entomologic study was conducted to study host preferences, insecticide susceptibility, anopheline diversity, seasonal variations, and intensity of malaria transmission (EIR). *Anopheles arabiensis* has developed resistance to the pyrethroid insecticides and DDT, and showed a greater tendency to feed on cattle than humans, with an overall human blood index (HBI; the proportion of the *An. arabiensis* fed on human blood meals of the total blood meals determined) of 44% and a bovine blood index of 69%. We report anopheline species diversity, monthly variations of *An. arabiensis* in relation to meteorologic variables, variations between houses in relation to distance from breeding sites, and malaria transmission indices of sporozoite rates and EIR.

**MATERIALS AND METHODS**

**Study area.** The study was conducted in Chano (Figure 1), which is north of Arba Minch during May 2009–April 2010.
The health post, which is at the center of the village, is located at 6°6.666′ N, 37°35.775′ E and at an altitude of 1,206 meters above sea level. For the purpose of epidemiologic study, the village was divided into three sub-villages and coded as sub-village 01, 02 and 03. A detailed description of the study area has been reported elsewhere.18

Monthly meteorologic data were recorded from the weather station at Arba Minch University, which is 6 km from the study area. In common with most areas in the southern Ethiopian Rift Valley,19 there were two wet seasons with the main rains falling during March–May and a smaller rainy season during October–December; the dry seasons are during June–September and January–February. Because of high surface water, we classified November 2009 as a wet month despite the low rainfall recorded.

Mosquito sampling. Thirty houses from the three sub-villages were selected for mosquito collections. The selected houses were distributed on the periphery and in the middle of each sub-village and equally divided for sampling by using CDC light traps (10 houses), PSCs (10 houses), or artificial pit shelters (10 houses). The distance from the main larval breeding sites to each house was recorded by using a global positioning system (GPS 60™; Garmon, Olathe, KS).

Anopheles mosquitoes were sampled every second week for 1 year (May 2009–April 2010). Before mosquito collection, verbal consent from the head of each household was obtained. Indoor host-seeking Anopheles were collected from 6:30 PM to 6:00 AM in 10 houses by using CDC light traps by hanging them 45 cm above the floor at the feet of sleeping persons, who were protected by untreated mosquito nets.13 Other occupants in the houses were left to use LLINs provided by the Ministry of Health as part of the routine malaria control. Two trained field assistants from the community turned the light traps on and off at 6:30 PM and 6:00 AM, respectively. In the mornings, collection bags were transported to the entomology laboratory at Arba Minch University for sorting and further processing.

Indoor-resting mosquitoes were sampled in the mornings (6:00 AM to 9:00 AM) from 10 other randomly selected houses by using the PSC technique. All food items and small animals were removed from houses, and all openings and eaves of windows and doors were closed with pieces of cloth. Finally, the floor and furniture were covered with white sheets before spraying houses with a pyrethroid roach killer aerosol (registration no. ET/HHP/130M/S, Kafr; EI Zayat, Egypt). Ten minutes after spraying, anophelines that had been knocked down were collected by using forceps, paper cups, and a torch light. The number of occupants who had slept in each house the previous night was recorded.

Outdoor-resting mosquitoes were collected by using a handheld mouth aspirator, paper cup, and torch from the pit shelters constructed under the shade of mango trees in the compound of each of the 10 houses. Each shelter was 1.5 m deep and had an opening of 1.2 m × 1.2 m. Approximately 0.5 meters from the bottom of each pit shelter, a 30-cm horizontal deep cavity was prepared in each of the four sides.20 During the collection period (6:30 AM–10:00 AM), the mouth of each pit shelter was covered with an untreated bed net to prevent mosquitoes from escaping.

Anopheles mosquito processing. Female Anopheles mosquitoes were killed by freezing, identified to the species level by using a morphologic key,21 and classified into unfed, freshly fed, half-gravid, and gravid.12 The abdomens of unfed An. arabiensis, the only member of the An. gambiae complex in the area,18 were dissected for parity based on the method of Detinova.22 The remaining parts of parous and other specimens of Anopheles were preserved individually in vials with silica gel for detection of circumsporozoite protein (CSP).

Detection of CSP. The head and thorax of female Anopheles were tested for the CSPs of Plasmodium falciparum,
P. vivax 210, and P. vivax 247 by enzyme-linked immunoabsorbent assay (ELISA). All positive samples were re-tested for confirmation.

Data and statistical analysis. Data were entered and analyzed by using SPSS version 19 (SPSS Inc., Chicago, IL). The sporozoite rate was calculated as the proportion of mosquitoes positive for P. falciparum and P. vivax of the total number of mosquitoes tested. Parity rate was determined as the proportion of parous mosquitoes over the number of total mosquitoes dissected.

Analysis of variance was used to compare the monthly total and the house density of An. arabiensis. Tukey’s honestly significant difference (HSD) test was used to distinguish the months and the houses with the maximum mean density of mosquitoes. Log-transformed data were used for statistical analysis. The Spearman’s rho correlation was used to test the relationship between mean monthly density of mosquitoes and the rainfall. All tests were performed at a 0.05 significance level.

The annual P. falciparum and P. vivax EIR of An. arabiensis was calculated from CDC light traps by using the standard method, 1,605 × (no. circumsporozoite-positive ELISA results from CDC light trap/no. mosquitoes tested) × (no. mosquitoes collected from CDC light trap/no. catches) × 365 days, and the alternative method, 1,605 (no. positive ELISA/no. catches) × 365 days. The monthly EIR was determined as a product of the EIRs for each day of the month. The EIR was also estimated from the PSC as described by the World Health Organization by using the formula: (human-biting rate) × (CSP rate). The human-biting rate was calculated by dividing the total number of freshly fed An. arabiensis caught by PSC by the total number of occupants who slept in the houses the night before collection and multiplied by the HBI. The HBI was calculated as the proportion of Anopheles mosquitoes that fed on humans (including mixed blood meal origins) of the total Anopheles analyzed for blood meal origin. Results of the HBI have been reported elsewhere.

RESULTS

Species composition. Overall, 4,708 anopheline mosquitoes belonging to 16 species were collected and identified. Of the 16 species collected, 14 species were obtained from CDC light traps (n = 2,506), 12 species from pit shelters (n = 1,678), and 9 species from PSCs (n = 524). Anopheles arabiensis accounted for 89% of the mosquitoes from CDC light traps, 83.4% from the PSCs, and 63.0% from pit shelter collections. The next most abundant species was An. marshalli, which accounted for 9.1% of mosquitoes caught in CDC light traps, 13% in PSCs, and 31.3% in pit shelters (Table 1). Using CDC light traps as a reference, we found that the catch ratio of PSCs to CDC light traps was 0.2 and that of pit shelters to CDC light traps was 0.67 for all anophelines. The figures for An. arabiensis were 0.2 for PSC and 0.47 for pit shelters, and the respective values for An. marshalli were 0.29 and 2.3. Further analyses were then performed on samples of An. arabiensis.

Monthly density of indoor-biting and indoor- and outdoor-resting mosquitoes. The monthly indoor and outdoor density of An. arabiensis in relation to rainfall is shown in Figure 2. Monthly density of indoor host-seeking An. arabiensis varied significantly (degrees of freedom [df] = 11, F = 6.0, P < 0.002). Collections peaked in April 2010 with 53.4/CDC light trap/night. In August 2009, no mosquitoes were found because of extremely low rainfall in June and July 2009. However, over most of the wet months (October, November, March, and April) the variation was not significant (P > 0.05, by HSD test).

The density of indoor-resting An. arabiensis also varied significantly (df = 11, F = 5.5, P = 0.003) with a maximum of 7.6/house/day in April 2010. Seasonal trends were also reflected for outdoor-resting density of An. arabiensis (df = 11, F = 8.1, P < 0.001), which had a maximum of 22/pit shelter/day and a minimum of 0/pit shelter/day.

The density of An. arabiensis was significantly associated with a one-month lag of rainfall in collections from CDC light traps (r = 0.81, P < 0.001), PSCs (r = 0.79, P = 0.002), and pit shelters (r = 0.63, P = 0.03). However, this association was not significant when An. arabiensis density was measured against rainfall in the month of collection or against a two-month lag.

![Figure 2. Monthly density of Anopheles arabiensis collected by three methods in Chano, southwestern Ethiopia. CDC = Centers for Disease Control.](image-url)
Household variations in indoor-biting and resting mosquitoes. The association between densities of *An. arabiensis* and distance from the major breeding sites is shown in Figure 3. Mosquitoes were more abundant in houses located near main breeding sites on the shore of Lake Abaya than in those further away. Density of indoor-biting *An. arabiensis* differed significantly between houses (df = 9, F = 16.3, P < 0.001). Of 2,230 *An. arabiensis* sampled by CDC light traps, 74.3% (1,657) were from the 30% of houses that were closest to larval breeding sites. The distance of the houses closest to the breeding sites was 1,350–1,570 meters, and the density of indoor-biting *An. arabiensis* in these houses ranged from 5.6/CDC light trap/house/night to 46.4/CDC light trap/house/night. In contrast, for houses further from the shore of the lake (2,350–2,600 meters), the density ranged from 0.4/CDC light trap/house/night to 2.8/CDC light trap/night.

For PSCs, a similar trend in variation of indoor-resting density of *An. arabiensis* (df = 9, F = 8.5, P < 0.001) was observed. The density of *An. arabiensis* ranged between 3.3/house/day and 6.5/house/day in those houses closest to the breeding sites (1,360–1,530 meters) but ranged between 0.0 and 2.2/house/day in those houses further from the shore of the lake (1,980–2,510 meters). No significant variation was observed among houses between 1,360 and 1,530 meters from the shore of the lake (P = 0.83, by HSD test). The density of *An. arabiensis* in pit shelters also varied (df = 9, F = 9.0, P < 0.001).

Abdominal conditions and parity rates. Abdominal conditions of *An. arabiensis* from different collection sites are shown in Figure 4. Freshly fed *An. arabiensis* were dominant (60.3% of 3,724) followed gravid and half-gravid (33.2%), and the percentage of unfed was low (6.4%). The ratio of freshly fed *An. arabiensis* was higher in the PSCs (81% of 437) and pit shelters (85.1% of 1,057) than in CDC light traps (44.5% of 2,230). The proportion of gravid (including half-gravid) to freshly fed *An. arabiensis* was 1.35 times higher in PSCs than in pit shelters.

Unfed *An. arabiensis* were collected almost exclusively in CDC light traps (98.3% of 239). Of the small number of unfed *An. arabiensis* caught and dissected (n = 239) for ovarian ageing, the parous rate was 26.4% (63 of 239). Sixty-two percent (149 of 239) of unfed and 68% (43 of 63) of parous *An. arabiensis* were collected from the house nearest to the shore of the lake.

Sporozoite rates. Overall, 4,534 *Anopheles* were analyzed for CSPs comprising *An. arabiensis* (n = 3,678), *An. marshalli* (n = 763), *An. garnhami* (n = 45), *An. funestus* group (n = 26), *An. pharoensis* (n = 15), and *An. tenebrosus* (n = 7). Of these mosquitoes, 14 *An. arabiensis* were positive for *Plasmodium* CSPs, 11 were positive for *P. falciparum* (78.6%), and 3 were positive for *P. vivax* 210 (21.4%). None of the tested *An. arabiensis* was positive for *P. vivax* 247 CSP, and no other anophelines were found to be positive for CSPs.

Monthly sporozoite rates of *An. arabiensis* from different collection sites are shown in Table 2. The greater numbers of (93% of 14) *Plasmodium*-positive *An. arabiensis* were collected during the wet months (October and November 2009 and March and April 2010). Seven of 11 *P. falciparum* sporozoite-positive *An. arabiensis* were captured by CDC light traps, three were collected in artificial pit shelters (3 of 11), and 1 by PSCs (1 of 11), although there was no statistically
significant difference between the collection methods. The number of *P. vivax*-positive *An. arabiensis* was similar from all collection sites. The overall *Plasmodium* infection rate (*P. falciparum* and *P. vivax*) of *An. arabiensis* was 0.38%, and the *P. falciparum* sporozoite rate was 0.32% for CDC light traps, 0.28% for pit shelters, and 0.23% for PSCs. When categorized by species, the overall rate was 0.3% for *P. falciparum* and 0.08% for *P. vivax* 210.

**Entomologic inoculation rates.** The monthly EIRs of *An. arabiensis* estimated from CDC light traps and PSCs are shown in Table 3. The monthly EIRs of *An. arabiensis* were highest in the wet months. A one-month lag of rainfall was significantly associated with the monthly EIR (*r* = 0.74, *P* = 0.006) of *An. arabiensis* from CDC light traps, but there was no significant association with the month of collection or with a two-month lag of rainfall (*P* > 0.05).

The estimated annual EIRs of *Anopheles* from CDC light traps and PSCs are shown in Table 4. Based on the alternative method, the estimated annual *P. falciparum* EIR of *An. arabiensis* from CDC light traps was 17.1 (95% confidence interval [CI] = 7.0–34.6 ib/p/year, whereas that of *P. vivax* was 2.4 (95% CI = 0.06–13.4). The EIR from PSCs was 0.1 ib/p/year for *P. falciparum* and *P. vivax*.

Estimates of the EIRs of *An. arabiensis* were also made individually for the three sub-villages, and for the wet (including main and small rainy months) and dry seasons for CDC light traps (Table 5). The *P. falciparum* EIR was 2.4 (95% CI = 0.12–11.7) in the dry season and 14.7 (95% CI = 5.9–29.4) in the wet season. This finding represented 6.1-fold more infectious bites in the wet than in the dry season. In sub-village 03, *P. falciparum* EIR was 24.4 ib/p/year (95% CI = 6.7–60.3) and *P. vivax* EIR was 5.8 ib/p/year (95% CI = 0.3–29.3). The annual *P. falciparum* EIR of *An. arabiensis* from CDC light traps varied between houses from 0 ib/p/year (in 60% of houses) to 73.2 (95% CI = 15.6–187) ib/p/year in house nearest to the major breeding site.

From the PSCs, two of the *An. arabiensis* collected on the same day from the house nearest the breeding site (with the maximum biting density of mosquitoes) were positive for CSP (1 for *P. falciparum* and 1 for *P. vivax*). The mean biting rate of *An. arabiensis* was 0.33 b/p/n, and the estimated EIR was 0.1 ib/p/year, which was calculated by multiplying the mean

### Table 2

Monthly circumsporozoite protein–positive *Anopheles arabiensis* collected by three methods from Chano in southwestern Ethiopia *

<table>
<thead>
<tr>
<th>Month and year</th>
<th>CDC light trap</th>
<th>PSC</th>
<th>Pit shelter</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>No. tested</td>
<td>% (95% CI)</td>
<td>No. tested</td>
</tr>
<tr>
<td>May 2009</td>
<td>73</td>
<td>0 (0.00)</td>
<td>90</td>
</tr>
<tr>
<td>Jun 2009</td>
<td>105</td>
<td>1 (0.95)</td>
<td>8</td>
</tr>
<tr>
<td>Jul 2009</td>
<td>8</td>
<td>0 (0.00)</td>
<td>1</td>
</tr>
<tr>
<td>Aug 2009</td>
<td>0</td>
<td>0 (0.00)</td>
<td>0</td>
</tr>
<tr>
<td>Sep 2009</td>
<td>28</td>
<td>0 (0.00)</td>
<td>6</td>
</tr>
<tr>
<td>Oct 2009</td>
<td>361</td>
<td>1 (0.76)</td>
<td>67</td>
</tr>
<tr>
<td>Nov 2009</td>
<td>356</td>
<td>1 (0.28)</td>
<td>67</td>
</tr>
<tr>
<td>Dec 2009</td>
<td>76</td>
<td>0 (0.00)</td>
<td>4</td>
</tr>
<tr>
<td>Jan 2010</td>
<td>87</td>
<td>0 (0.00)</td>
<td>14</td>
</tr>
<tr>
<td>Feb 2010</td>
<td>33</td>
<td>0 (0.00)</td>
<td>3</td>
</tr>
<tr>
<td>Mar 2010</td>
<td>224</td>
<td>0 (0.00)</td>
<td>24</td>
</tr>
<tr>
<td>Apr 2010</td>
<td>1,063</td>
<td>4 (0.38)</td>
<td>150</td>
</tr>
<tr>
<td>Total</td>
<td>2,184</td>
<td>7 (0.32)</td>
<td>436</td>
</tr>
</tbody>
</table>

* CDC = Centers for Disease Control; PSC = pyrethrum spray catch; Pl = *Plasmodium falciparum*; Cl = confidence interval; Pv = *P. vivax*.

### Table 3

Monthly EIR of *Anopheles arabiensis* from CDC light traps and pyrethrum spray catches in Chano, southwestern Ethiopia *

<table>
<thead>
<tr>
<th>Month and year</th>
<th>CDC</th>
<th>PSC</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>No. tested</td>
<td>No. tested for CSP</td>
</tr>
<tr>
<td>May 2009</td>
<td>76</td>
<td>73</td>
</tr>
<tr>
<td>June 2009</td>
<td>114</td>
<td>105</td>
</tr>
<tr>
<td>July 2009</td>
<td>8</td>
<td>8</td>
</tr>
<tr>
<td>August 2009</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>September 2009</td>
<td>33</td>
<td>28</td>
</tr>
<tr>
<td>October 2009</td>
<td>140</td>
<td>131</td>
</tr>
<tr>
<td>November 2009</td>
<td>361</td>
<td>356</td>
</tr>
<tr>
<td>December 2009</td>
<td>77</td>
<td>76</td>
</tr>
<tr>
<td>January 2010</td>
<td>90</td>
<td>87</td>
</tr>
<tr>
<td>February 2010</td>
<td>36</td>
<td>33</td>
</tr>
<tr>
<td>March 2010</td>
<td>226</td>
<td>224</td>
</tr>
<tr>
<td>April 2010</td>
<td>1,069</td>
<td>1,063</td>
</tr>
</tbody>
</table>

* EIR = entomologic inoculation rate; CDC = Centers for Disease Control; PSC = pyrethrum spray catch; CSP = circumsporozoite protein; CI = confidence interval; Pl = *Plasmodium falciparum*; Pv = *P. vivax*.

† Standard method: 1.605 (no. enzyme-linked immunosorbent assay [ELISA] positive from CDC light trap/no. ELISA tested) × (no. *An. arabiensis* collected from CDC light trap/no. of catches) × no. days per month.

‡ Alternative method: 1.605 (no. ELISA positive/no. catches) × no. days per month.

§ *P. vivax* EIR.
human-biting density by HBI (from a previous report\textsuperscript{18}) and the CSP rate.

**DISCUSSION**

This study showed that the estimated annual *P. falciparum* EIR of *An. arabiensis* in Chano, Ethiopia was 17.1 infectious bites/person/year. *Anopheles arabiensis* was identified as the principal vector of *Plasmodium* in the area, and the risk of exposure to infectious bites was higher in wet seasons than in dry seasons. There was a high variation in EIRs between houses, and a maximum in the house closest to larval-breeding sites.

The estimated EIR from CDC light traps was higher and relatively more representative than that of PSCs because a greater number of *Plasmodium*-positive *An. arabiensis* were collected in CDC light traps. We concluded that the EIR from PSCs cannot be representative of the study area because the two CSP-positive specimens were sampled from one house on the same day. The PSCs also had lower relative sampling efficiency for *Anopheles* mosquitoes than CDC light traps. The most likely explanation is that indoor-resting adults might leave houses immediately after feeding\textsuperscript{7} before spraying, and during spraying.\textsuperscript{15} Therefore, using PSCs for estimating EIR can underestimate the human-biting rates.\textsuperscript{15}

Anopheles arabiensis was the principal vector of malaria in the study area, as reported.\textsuperscript{2,14} We identified 16 species of anophelines, of which 5 had been reported from southern Ethiopia.\textsuperscript{14} Unlike most studies in this country,\textsuperscript{2,14,30} *An. pharoensis* were rarely sampled in our study site. *Anopheles marshalli* was the second most abundant species although in a previous study from Sille in southern Ethiopia, it was found only at low densities.\textsuperscript{14} The high proportion of HBI\textsuperscript{18} and the moderately frequent occurrence of *An. marshallii* indoors suggests a need for further investigations into its potential role as a vector of malaria in this area and other areas in Ethiopia.

The parous rate of *An. arabiensis* in our study was lower than the 73.2\% reported in Sille in 2006\textsuperscript{14} and the 58.3\% reported in Awash Valley in 1996.\textsuperscript{31} This difference could be the result of mass emergence of nulliparous adults from the nearest breeding sites\textsuperscript{12} because most unfed *An. arabiensis* were collected from a house near the shore of the lake. The proportion of gravid (including half-gravid) to freshly fed *An. arabiensis* was 1.35 times higher for PSCs than for pit shelters, which suggested a tendency for endophilic rather than exophilic behavior. Exophilic and endophilic behavior of *An. arabiensis* has been reported in southern Ethiopia.\textsuperscript{18,33}

We observed a clustered distribution of *Plasmodium* CSP-positive *An. arabiensis* in a sub-village near the shore of the lake. This finding is consistent with recently reported clustering of malaria cases from the same part of a village\textsuperscript{34} and the

**Table 4**

<table>
<thead>
<tr>
<th>Variable</th>
<th>CDC light trap</th>
<th>PSC</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>No. collected</td>
<td>No. positive/no. tested (%)</td>
</tr>
<tr>
<td><em>Pf</em> EIR</td>
<td>2,230</td>
<td>7/2,184 (0.32)</td>
</tr>
<tr>
<td><em>Pv</em> EIR</td>
<td>2,230</td>
<td>1/2,184 (0.046)</td>
</tr>
</tbody>
</table>

\textsuperscript{†}Alternative method: 1.605 (no. enzyme-linked immunosorbent assay [ELISA] positive from CDC light trap/no. ELISA tested) \textsuperscript{‡}Alternative method: 1.605 (no. ELISA positive/no. catches) \textsuperscript{§}EIR calculated by multiplying by 182 days. \textsuperscript{¶}EIR calculated by multiplying by 183 days.

**Table 5**

<table>
<thead>
<tr>
<th>Variable</th>
<th>No. collected</th>
<th>No. tested</th>
<th>No. catches</th>
<th>No. positive (%)</th>
<th><em>Pf</em> EIR†</th>
<th><em>Pf</em> EIR† (95% CI)</th>
<th>No. positive (%)</th>
<th><em>Pv</em> EIR†</th>
<th><em>Pv</em> EIR† (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sub-village 01</td>
<td>124</td>
<td>117</td>
<td>72</td>
<td>0 (0.0)</td>
<td>0.0</td>
<td>0.0 (0.0–24.0)</td>
<td>0</td>
<td>0.0</td>
<td>0.0 (0.0–24.0)</td>
</tr>
<tr>
<td>02</td>
<td>411</td>
<td>409</td>
<td>72</td>
<td>3 (0.73)</td>
<td>24.5</td>
<td>24.4 (51.6–68.5)</td>
<td>0</td>
<td>0.0</td>
<td>0.0 (0.0–24.0)</td>
</tr>
<tr>
<td>03</td>
<td>1,695</td>
<td>1,658</td>
<td>96</td>
<td>4 (0.24)</td>
<td>25.0</td>
<td>24.4 (67.6–60.3)</td>
<td>1 (0.06)</td>
<td>6.2</td>
<td>5.8 (3.2–29.5)</td>
</tr>
<tr>
<td>Season Wet</td>
<td>1,949</td>
<td>1,923</td>
<td>120</td>
<td>6 (0.31)</td>
<td>14.9</td>
<td>14.7 (5.9–29.4)</td>
<td>1 (0.05)</td>
<td>2.5</td>
<td>2.4 (12.117)</td>
</tr>
<tr>
<td>Dry</td>
<td>281</td>
<td>261</td>
<td>120</td>
<td>1 (0.38)</td>
<td>2.6</td>
<td>2.4 (12.117)</td>
<td>0</td>
<td>0.0</td>
<td>0.0 (14.6)</td>
</tr>
</tbody>
</table>

\textsuperscript{*}Alternative method: 1.605 (no. enzyme-linked immunosorbent assay [ELISA] positive from CDC light trap/no. ELISA tested) \textsuperscript{†}Alternative method: 1.605 (no. ELISA positive/no. catches) \textsuperscript{‡}EIR calculated by multiplying by 182 days. \textsuperscript{§}EIR calculated by multiplying by 183 days.
distribution of malaria vectors in Sille in southern Ethiopia. The *P. falciparum* sporozoite rate of *An. arabiensis* from CDC light traps (0.32%) is comparable with that reported from Sille (0.5%) for HLCs, but the overall sporozoite rate (0.38%) was lower than the 2.26% rate in Sille. A higher *P. falciparum* CSP rate of *An. arabiensis* from CDC light traps was also reported from Ziway in the Central Rift Valley of Ethiopia (1.18%). Krafsur in 1977 reported a higher sporozoite rate (1.87%) for indoor-resting *An. arabiensis*, and in a report in 2013 for south-central Ethiopia showed a *P. falciparum* CSP rate of 0.3% for CDC light traps and 0.2% for PSCs for *An. arabiensis*.

This study showed that 93% of sporozoite-positive *An. arabiensis* were found in the wet season and together with EIR were closely associated with rainfall, as has been demonstrated in Gambela in western Ethiopia, in Ifakara, Tanzania, in Eritrea and elsewhere in Africa. It is clear that the risk of malaria transmission increases during periods of higher EIR and a decrease in density and number of sporozoite-positive mosquitoes results in a decreased EIR. Based on CDC light traps, the estimated annual *P. falciparum* EIR (17.1 ib/p/year) of *An. arabiensis* was higher compared with EIRs of *An. arabiensis* in Gambela (5.36 ib/p/year) and nearby villages (10.47 ib/p/year) estimated from PSCs but much lower than the overall EIR from river villages in Gambela (96.67 ib/p/year). Recently, an annual *P. falciparum* EIR of 3.66 ib/p/year for *An. arabiensis* was reported from the central highland of southern Ethiopia. However, in two other studies in neighboring countries, higher EIRs than those recorded in our study have been reported. One study that lasted more than two years and used HLCs reported EIRs of 70.6 ib/p/year from Hiletsidi in the Gash Barka zone and 32.1 ib/p/year in Maiaini in the Debub zone in Eritrea, and another study using the PSC method estimated an annual EIR for *An. arabiensis* of 55.48 ib/p/year in eastern Sudan.

The EIR has implications for monitoring the suitability of vector control interventions. Current malaria vector interventions such as LLINs and IRS reduce EIR in many malaria-endemic countries, but none of these interventions has managed to reduce EIR to < 1 ib/p/year, except in the Garki Project (which used propoxur for IRS) and reduced EIR only temporarily. It has been suggested that the annual EIR should be < 1 ib/p/year to reduce and interrupt malaria transmission, based on the finding of a linear relationship between malaria prevalence and EIR. Our EIR estimate (17.1 ib/p/year) is more than sufficient to continue malaria transmission in the area, where incidence is reported to be 3.6/10,000 person-weeks of observations.

Finally, this study clearly identified *An. arabiensis* as the principal vector of malaria in the Chano area, and the estimated EIR from CDC light traps was higher and more representative than that of PSCs. The risk of exposure to infectious bites was higher in the houses closer to the larval breeding sites, and in wet seasons than in dry seasons. Besides advocating about using the malaria vector control programs for the general population, we advise the vector control programs to focus those households with malaria clustering (hot spots). Such interventions could include IRS during the seasons of the local malaria vector density. Currently, we study if combining screening doors and windows with IRS and LLINs for those houses closer to the breeding sites will reduce malaria transmission. Because the nearby lake was the main mosquito breeding site, it might be worthwhile to assess if aerial spraying of larvicide on the lakeshore would reduce malaria in such populations.

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REFERENCES


