Case Report: Transient Success Using Prolonged Treatment with Miltefosine for a Patient with Diffuse Cutaneous Leishmaniasis Infected with Leishmania mexicana mexicana

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Abstract. Leishmania sp. is an intracellular parasite that causes a variable degree of clinical manifestations, especially in the skin. We present the case of a 38-year-old male with a chronic history of mucocutaneous disease present since childhood that generated deformity, loss of cartilage in the ears and nose, and scarring that limited his range of motion. The parasite was identified as L. mexicana mexicana. The patient was treated with a 3-month course of oral miltefosine with overwhelming results.

INTRODUCTION

Leishmaniasis is a vector-borne disease caused by obligate intracellular protozoa of the genus Leishmania found in the tropics that is transmitted by the Phlebotomine sandflies.1

There are different clinical presentations of the disease that include visceral, mucosal, and cutaneous, but the latter is the most common.2 Approximately 21 species of Leishmania can infect humans. These species include the L. donovani complex, L. mexicana complex, L. tropica, L. major, L. aethiopica, and the subgenus Viannia. Species of Leishmania are morphologically indistinguishable by conventional methods, but they can be differentiated by isoenzyme analysis, molecular methods, or monoclonal antibodies. The anergic or diffuse disease in Mexico is very rare, especially in immunocompetent patients without a travel history to endemic areas. Standard treatment of leishmaniasis is based on antimonials, but a variety of drugs are being used in patients with mucocutaneous disease with promising results.3 4 Miltefosine is a phosphocholine analog that is administered orally; although originally used in visceral disease, it has been used for cutaneous disease with variable results.

We present a case of mucocutaneous disease caused by L. mexicana in a non-immunocompromised Mexican patient without travel history that was refractory to combined treatment and responded to miltefosine.

CASE PRESENTATION

A 38-year-old man, a native and resident of Mendez County, Tamaulipas (93.7 miles south of the Mexico–Texas border), with no travel history outside the region presented to the emergency room with stridor.

He had a history of anergic leishmaniasis since his childhood and had been treated during that time with inconsistent results. The patient had been lost to follow-up during the previous 15 years and was in his usual state of health until the previous 6 months, when he noticed vocal changes with reduced tone and volume. He developed progressive dysphagia and malaise and subsequently, lost 10 kg body weight.

Three months before arrival, he noticed progressive dyspnea and stridor. The latter became more intense, and he finally sought medical attention.

He was transferred to our hospital from his local medical clinic for definitive treatment. His blood pressure was 110/70 mmHg, pulse was 80 bpm, respiratory rate was 16 breaths per minute, axillary temperature was 36°C, and pulseoximetry was 98% while on room air. The patient had a disseminated asymmetrical polymorphic dermatosis characterized by multiple non-tender nodules of 1–7 cm in diameter. The nodules were polypoid and pink/flesh-colored with an elastic consistency; some were superficial, and some were fixed to deep tissue (Figure 1A). Cartilage destruction was present in both ears and nose. During direct laryngoscopy, the oropharyngeal mucosa had a pavement aspect, and granulomatous lesions were observed in the soft palate and extended to epiglottis and glottis. Severe laryngeal stenosis was documented, and an emergency tracheostomy was performed. The rest of the physical exam was unremarkable.

Supraglottic biopsies showed amastigotes compatible with leishmaniasis associated with papillomatous changes. Various skin biopsies also showed amastigotes from Leishmania spp. The species was subsequently identified as Leishmania mexicana by polymerase chain reaction (PCR).

Blood count, serum electrolytes, and liver and kidney function tests were normal. A chest X-ray, electrocardiogram (EKG), and abdominal ultrasound were all normal. The patient is human immunodeficiency virus (HIV) -negative on repeated test. Lymphocytes CD3 count was 1,103 cells/mL (68.6%), lymphocytes CD3/CD4 count was 567 cells/mL (35.2%), lymphocytes CD3/CD8 count was 593 cells/mL (36.8%), natural killer cells count was 51.3 cells/mL (3.1%), lymphocytes CD19 count was 129 (8.0%), immunoglobulin A (IgA) count was 14.7 mg%, IgE count was 8.3 mg%, IgG count was 727.4 mg%, IgM count was 22.9 mg%, complement C3 count was 57.1 mg%, complement C4 count was 17.6 mg%, and CH50 count was 8.3 mg% (Table 1).

Treatment was initiated with intravenous (IV) meglumine (20 mg/kg per day), omeprazole per Ora (PO) (20 mg/day), and IV amphotericin B (0.5–0.75 mg/kg per day). Gradual improvement was observed on day 28 of treatment with no toxicity, but the lesions had not disappeared (Figure 1B). We extended the treatment to 38 days, with a total of 38 doses of meglumine and a cumulative dose of amphotericin B of 1,145 mg. Repeated EKGS and liver and renal screens were normal.
The patient requested voluntary discharge, and by 60 days follow-up, although no new lesions were noticed, the previous lesions had not disappeared completely. He was decanulated after glottis plasty without complications.

The patient returned after 5 months with new lesions on his extremities and eyelids (Figure 1B). We started treatment with 100 mg miltefosine (50 mg bis in die [BID] PO). An early and marked improvement was observed with miltefosine. Treatment was continued for 3 months. No toxicity was documented (Figure 1C). Unfortunately, 6 months after the patient stopped miltefosine treatment, the lesions gradually reappeared with their usual distribution and characteristics (Figure 1D).

**DISCUSSION**

The clinical spectrum of cutaneous leishmaniasis in humans includes localized cutaneous leishmaniasis, mucocutaneous leishmaniasis, disseminated cutaneous leishmaniasis, and diffuse cutaneous leishmaniasis. The clinical manifestations are determined by the characteristics and the immune response of the infected patient.5

Localized cutaneous leishmaniasis is characterized by a single or multiple erythematous papules, usually located in the exposed region, that develop into ulcers with raised borders. The involvement of the nasal mucosa, palate, pharynx, larynx, and vocal cords may occur in up to 5% of the patients.

Diffuse cutaneous leishmaniasis (DCL) is a rare variant that usually begins in infancy and is characterized by the appearance of erythematous or skin-colored papules, plaques, or nodules with a continuous and uncontrolled dissemination; in advanced stages, there may be a parasitic invasion to nasopharyngeal and oral mucosa in more than 30% of the patients.6

It is necessary to establish the clinical difference with the disseminated cutaneous leishmaniasis, which is characterized by the presence of multiple polymorphic lesions that may present as multiple papules and acneiform-type lesions with few

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$\text{CH50} = \text{complement hemolyzing 50}; \text{UCH} = \text{units of complement hemolyzing 50}.$
ulcers and mucosal involvement in 38% of cases. It is accom-
panied by symptoms such as fever and malaise. The immuno-
ological aspects are also distinct, because there is an abnormality in
 cellular immunity that is restored after treatment.7

For the pathogenesis of anergic DCL, there is a deficiency in
 cellular immune response that leads to poor parasite clear-
 ance and chronicity of the disease. It is found that there is an
 alteration in the cellular response T-helper (Th)-2 type, which is
 associated with a specific cell anergy to leishmania antigens
 and manifested by a negative Montenegro reaction.8 Also,
 there is a low level of CD-8 T lymphocytes, which plays an
 important role against infection by being a potential target for
 a vaccine development.9

Important for development of immunotherapy agents, they
can exert their effect by directly or indirectly augmenting the
 host natural defenses, thereby restoring the impaired effector
 functions or decreasing host excessive and deleterious
 response to the parasite. Initially, dead promastigotes were
 used for the development of prophylactic vaccines in Brazilian
 patients.10 The work by Pereira and others11 reported the use
 of a vaccine containing killed promastigotes of Leishmania
 associated with the Bacillus Calmette–Guerin (BCG) alone
 or combined with chemotherapy. This vaccine may be an al-
 ternative for patients who are intolerant or resistant to conven-
tional treatment. The new vaccines are based on highly purified
 biological compounds and cytokines, interleukin-2 (IL-2), IL-4,
 IL-12, and interferon (INF-γ) which have the specific function
 of regulating the cellular response of Th-1and Th-2.12

DCL is associated with a poorer prognosis; the most chal-
 lenging issue is the refractory nature of the condition to treat-
 ment, making it a demanding and often frustrating task.

This case illustrates the difficulty in obtaining sustained
 response to treatment in diffuse cutaneous leishmaniasis. This
 presentation of the disease is very rare in Mexico.

To our knowledge, no other cases similar to this case have
 been reported in the northern region of the country during
 the last 40 years.13 L. mexicana has been well-documented in
 Texas, and it causes human and animal infections, with the
 majority being single cutaneous lesions.14–16

This case was first treated by combining meglumine and
 amphotericin B (both medications are commonly used as
 monotherapy) in addition to using omeprazole, which induces
 changes in the acid pH that favors the proliferation of
 amastigotes in macrophages17; combination with antiparasitic
 drugs has been described mainly in visceral leishmaniasis
 (VL), and some studies are showing encouraging results.18–20

In this patient, the combination was successful as initial treat-
 ment without evidence of potentially toxic effects after 5 weeks
 of treatment. Although cardiovascular, liver, and kidney toxic-
 ity have been described with both medications,2,20 in our
 patient, strict cardiovascular monitoring did not show any
 abnormalities during and after combination therapy.

Miltéfosine has shown a cure rate in immunocompetent
 patients with VL of 94% in India and about 90% in Ethiopia,
 where L. donovani is the cause of VL; it is listed as an option for
 VL in geographic areas where L. donovani is the predominant
 cause of VL.2 For geographical areas where L. infantum is the
 cause of VL (examples: Mediterranean Basin, Middle East, Cen-
 tral Asia, and South America), miltéfosine is not recommended.2

In a non-randomized trial using miltéfosine (2.5 mg/kg per
day for 28 days) for L. braziliensis, the cure rate for the
36 patients who had extensive disease (involving the palate,
 pharynx, and larynx) was 58%. For the contemporary group
that was receiving amphotericin B (45 mg/kg over 90 days),
the cure rate was 7 (50%) of 14 patients.21 We decided to start
 treatment with miltéfosine and extend it for 3 months based
on the encouraging results.

Relapse after miltéfosine treatment has been described pre-
viously22–23 in a patient with diffuse cutaneous leishmaniasis
caused by L. mexicana. Our patient followed a similar relapse
pattern, although the time free of new lesions in our patient
was longer (6 versus 2 months). The treatment of DCL can be
very frustrating, because there is no sustained response to
 treatment. In our case, the response to a combined therapy
with amphotericin B and meglumine was transitory like the
response to miltéfosine. In our case, the advantage with
miltéfosine is the outpatient setting and the possibility of
extended treatment.

CONCLUSION

Mucocutaneous leishmaniasis caused by L. mexicana clini-
cally responds to treatment with miltéfosine with a high proba-
bility of relapse. Duration of treatment beyond 28 days seems
to be well-tolerated with no toxicity. This finding suggests that
miltéfosine could be a useful maintenance therapy in DCL in
an outpatient setting. Because of the immune deficit and phys-
 iopathology of DCL, a curative treatment is still not available.

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REFERENCES

1. Maroli M, Khoury C. 2004. Prevention and control of leishmani-
2. World Health Organization. Control of Leishmaniasis. Report of
a Meeting of the WHO Expert Committee on the Control of
Leishmaniasis. Available at: http://whqlibdoc.who.int/trs/TRS
 treatment of visceral leishmaniasis. Expert Rev Anti Infect Ther
8: 919–944.
4. Richard JV, Werbovetz KA. 2010. New antileishmanial candi-
5. Almeida OL, Santos JB. 2011. Advances in the treatment of
 cutaneous leishmaniasis in the new world in the last ten years:
6. Zerpa O, Ulrich M, Blanco B, Polegren M, Avila A, Matos N,
Mendoza I, Pratlong F, Ravel C, Convit J, 2007. Diffuse cuta-
necous leishmaniasis responds to miltéfosine but then relapses.
7. Turetz ML, Machado PR, Ko AL, Alves F, Bittencourt A,
Almeida RF, Mobashery N, Johnson WD Jr, Carvalho EM,
2002. Disseminated leishmaniasis: a new and emerging form
of leishmaniasis observed in northeastern Brazil. J Infect Dis
186: 1829–1834.


