Usefulness of the rK39-Immunochromatographic Test, Direct Agglutination Test, and Leishmanin Skin Test for Detecting Asymptomatic Leishmania Infection in Children in a New Visceral Leishmaniasis Focus in Amhara State, Ethiopia

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Abstract. In areas where visceral leishmaniasis is anthropoontic, asymptotically infected patients may play a role in transmission. Additionally, the number of asymptomatic patients in a disease-endemic area will also provide information on transmission dynamics. Libo Kemkem and Fogera districts (Amhara State, Ethiopia) are now considered newly established areas to which visceral leishmaniasis is endemic. In selected villages in these districts, we conducted a study to assess the usefulness of different approaches to estimate the asymptomatic infection rate. Of 605 participants, the rK39 immunochromatographic test was able to detect asymptomatic infection in 1.5% (9 of 605), direct agglutination test in 5.3% (32 of 605), and leishmanin skin test in 5.6% (33 of 589); the combined use of serologic methods and leishmanin skin test enabled detecting asymptomatic infection in 10.1% (61 of 605). We conclude that the best option to detect asymptomatic infection in this new visceral leishmaniasis–endemic focus is the combined use of the direct agglutination test and the leishmanin skin test.

INTRODUCTION

Visceral leishmaniasis (VL) is a vector-borne disease caused by members of the Leishmania (Leishmania) donovani complex. It is endemic to 65 countries and, among them, Bangladesh, India, Nepal, Brazil, Sudan, and Ethiopia account for approximately 90% of the cases. The estimated annual incidence is 500,000 clinical cases with 59,000 associated deaths.1,2 Poor populations are particularly affected by VL, which is considered as one of the most neglected diseases.3 In addition, VL is currently spreading and re-emerging in different areas of the world with increasing public health concern.4

Visceral leishmaniasis is fatal if left untreated, and even with treatment the case fatality rate ranges from 4% to 10%.5,6 In stable disease-endemic areas, clinical disease appears only in a fraction of those infected, and another fraction will not develop the disease and remain asymptomatic.7 The prevalence of asymptomatic infection is different between and within different disease-endemic countries, and the number of asymptomatic infections usually exceeds the number of symptomatic infections, although this ratio can vary from 0.4:1 to 50:1.2

Visualization of parasite amastigotes by microscopic examination of bone marrow, spleen, or lymph node aspirates has been the gold standard method of VL diagnosis for many years. However, because this procedure is based on invasive sampling, its use for asymptomatic infection surveillance is not justified. In addition, in poor, remote, disease-endemic areas, the expertise and facilities required for these procedures may not be present. Thus, procedures based on less invasive sampling, such as serologic analysis or leishmanin skin test (LST) seem to be more suitable for this purpose.

Although detection of antibodies against Leishmania does not discriminate between current or past infection, serologic methods have been used to assess asymptomatic infection in different VL-endemic areas. These methods have been based on the direct agglutination test (DAT), rK39-immunochromatographic test (rK39-ICT), enzyme-linked immunosorbent assay, indirect immunofluorescent antibody test, or Western blot.3,4,8-12

The LST is a useful method for detecting cell-mediated immunity against Leishmania. Results for this test become positive after subclinical infection and, in this case, persist much longer than antibodies against Leishmania. The LST results also become positive within weeks-months after successful therapy against VL, indicating a healing or protective response.13,14 This makes the LST a valuable tool in detecting exposure to Leishmania parasites in epidemiologic surveys, and its usefulness to detect asymptomatic infection has been shown by different authors in different disease-endemic areas.11,15,16 In general, the LST would detect a higher proportion of asymptomatic infection than serologic analysis. However, given that serologic analysis and LST are based on different types of immune responses, in the absence of a gold standard, the combination of these two approaches would give a more realistic picture of the asymptomatic infection rate in a given disease-endemic area.17,18

In areas where VL transmission is anthropoontic, asymptomatic persons might play a role as reservoirs, and even in areas where VL is zoonoctic it is speculated that these persons could also contribute to transmission.7,19 Thus, assessment of the prevalence and distribution of asymptomatic cases would contribute to a better understanding of VL transmission and help in developing control efforts.

In Ethiopia, VL is an endemic disease of increasing public health concern. It is estimated that 30% of VL patients in Ethiopia are malnourished and co-infection with human immunodeficiency virus (HIV) affects 40% in the northwestern region of this country, and these conditions are known to facilitate the spread of VL.20,21 In addition to the classical foci in

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the northwestern region along the border with Sudan (Humera and Metema) and those in the southern region (Lake Abaya region, Omo River, and Aba Roba plains), the disease has recently spread to previously non-endemic areas, such as Libo Kemkem and Fogera (highland districts in Amhara State) where a VL outbreak occurred in 2004–2005. Thus, VL is currently a priority in the public health agenda of the Amhara State Health Bureau, and there is a need to generate epidemiologic data on VL in Amhara State.

To support VL control, facilities for treatment, mobile teams for surveillance, community mobilization, and active case detection strategies have been established. To contribute to this initiative the UBS-Optimus Foundation supported the project entitled Visceral Leishmaniasis and Malnutrition in Amhara State, Ethiopia, which among its specific objectives aimed to characterize nutritional, immunologic, and parasitologic aspects in the child population from this area, and it is within the framework of this project that we have explored the usefulness of rK39-ICT, DAT, and LST to detect asymptomatic Leishmania infection in children from different sub-districts of Libo Kemkem and Fogera. Furthermore, detection of asymptomatic infection in children will provide information about the status of VL transmission in this highland focus. In addition, this information will contribute to assessment of the magnitude of asymptomatic infection, which can help in early detection and treatment, thus contributing to decreased transmission, morbidity, and mortality. The information obtained can also be useful in a scenario in which HIV spreads to Leishmania-endemic areas, and help to foresee new VL cases among asymptomatic persons as a consequence of a reactivation of previous Leishmania infections after acquired HIV.

**Materials and Methods**

**Study Site.** The study was conducted during May–July 2009 in the districts (weredas) of Libo Kemkem and Fogera, Amhara State, Ethiopia (Figures 1 and 2). These are adjacent districts most affected by the outbreak of VL that occurred in 2004–2005. According to the 2009 census, the population was 198,374 (males = 100,951 and females = 198,374) and 226,595 (males = 115,693 and females = 110,902) for Libo Kemkem and Fogera, respectively. The districts are located in a black cotton clay soil flat plain (1800–2000 meters above sea level). Human activities related to intensive cultivation of teff, maize, beans, oilseeds, rice, and cotton have reduced the natural vegetation to scattered clumps of acacia trees. Most of the area is flooded during the rainy season (July–September) and dried up during the dry season (November–May), resulting in deep cracks in the soil surface, which could become breeding sites for the putative vector *Phlebotomus orientalis*.

**Study Population.** Population sampling was carried out by using a multi-staged cluster survey. Primary sampling units were sub-districts (kebeles) with high incidence of VL according to the 2008 register of the Addis Zemen VL Treatment Center: Agita, Bura, and Yifag from Libo Kemkem district and Dibasifatra and Rib Gebriel from Fogera district. Secondary sampling units were randomly selected villages (gotts) in each of the selected sub-districts. Tertiary sampling units were randomly selected households in each of the villages. All children with a reported age between 4 and 15 years living in the household at the time of the survey were eligible for the study, as long as they were asymptomatic and had no history of VL.

![Location of the study area in Ethiopia.](image-url)
**Data collection.** Information on age, sex, residence, and clinical assessment was obtained for all participants by trained medical personnel (nurses and health officers) by using pretested questionnaires and protocols.

**Asymptomatic case definition.** Asymptomatic persons were those who had a positive result in the rK39-ICT, DAT, or LST, and the absence of VL signs and symptoms (fever for > 2 weeks, in combination with either enlargement of spleen and/or liver, or weight loss).

**Sample collection and storage.** Peripheral blood was collected in tubes containing Na₂-EDTA (Sigma, St. Louis, MO) and immediately one drop was used for the rK39-ICT and two drops were spotted onto Whatman 3MM filter paper (Whatman International Ltd., Maidstone, United Kingdom). Filter papers were left to air dry and placed individually in sealed plastic bags. The plastic bags containing filter papers were kept in a chilled icebox and sent on the same day to the Amhara State Regional Laboratory, where they were stored at 4°C for further DAT analysis.

**Ethical considerations.** The study was approved by the ethical review boards of Instituto de Salud Carlos III, the Armauer Hansen Research Institute, and the Ethiopian National Ethical Review Committee. Support letters were obtained from the Amhara State and health bureaus in different districts. Parents/guardians gave written informed consent before enrollment of their children in the study. For children > 11 years of age, verbal assents were also obtained in addition to the consent of their parents/guardians.

**Detection of antibodies against Leishmania.** The rK39-ICT (Kalazar Detect® Rapid Test; InBios International Inc., Seattle, WA) was performed by using 1 drop of blood and 3 drops of chasing buffer according to the manufacturer's instructions. The DAT with freeze-dried antigen (ITMA-DAT/VL; Prince Leopold Institute of Tropical Medicine, Antwerp, Belgium)
was performed by using the screening method according to the manufacturer's protocol. Blood samples with a titer ≥ 1:3,200 were considered positive.

**Leishmanin skin test.** The LST was performed by using *L. major* antigen (Leishmanin batch 123-2; Pasteur Institute, Tehran, Iran). One hundred microliters of the antigen were intradermally inoculated on the volar surface of the forearm with a 1-mL sterile syringe and disposable needle. The test was read 48 hours later by using the ballpoint pen method. An induration with an average of two perpendiculars ≥ 5 mm was considered positive.

**Data analysis.** Infection prevalence was calculated by using rK39-ICT, DAT, and LST results. The differences in infection prevalence between age group, sex, and location were compared by using Fisher's exact and chi-square tests. A *P* value < 0.05 was considered statistically significant. Data analysis was performed by using SPSS version 16.0 (SPSS Inc., Chicago, IL) and STATA version 10 (Statacorp LP, College Station, TX).

### RESULTS

A total of 639 children were screened; one of them was an active case of malaria, three had an active VL episode, and 28 reported having VL in the past. For two other children, their previous VL history was unknown. These findings resulted in 605 asymptomatic children without previous VL eligible for analyses. The active malaria and VL cases were referred to the Addis Zemen Health Center for further clinical examination and treatment.

Of the 605 participants, 309 were boys (51.1%) and 296 were girls (48.9%). The boy:girl ratio was approximately 1:1 in the two districts studied. The mean ± SD age of the participants was 8.8 ± 0.13 years (there was no difference between the sexes). The children were grouped according to their age into three groups: group 1 (< 5 years of age) consisted of 77 of 605 (12.7%) children, group 2 (5–9 years of age) consisted of 295 (48.8%) of 605 children, and group 3 (10–15 years of age) consisted of 233 (38.5%) of 605 children.

Antibodies against *Leishmania* were detected in 9 (1.5%) of 605 children by the rK39-ICT and in 32 (5.3%) of 605 by the DAT. Sixteen (2.6%) of the 605 children initially tested by LST were lost for analysis. This test showed a positive result for 33 (5.6%) of 589 children.

Globally, 38 (6.3%) of 605 children were seropositive (rK39-ICT and/or DAT positive), and 61 (10.1%) of 605 children were considered infected (positive by rK39-ICT and/or DAT and/or LST). For the group of 589 children tested by using the three methods, we observed that 27 of 37 seropositive children had a negative LST result, although 23 of 33 LST-positive children were seronegative. A detailed description of the performance of these three methods in the group of 589 children is shown in Table 1.

The mean ± SD age of those with asymptomatic infection was 10.6 ± 3.4 years. Analysis by age group showed a positive association between asymptomatic infection and age, and children 10–15 years of age had a higher asymptomatic infection rate (16.3%; *P* < 0.0001). This finding was common for all tests used.

A strong association was also found with regard to sex, with infection in boys being higher than in girls (13.9% versus 6.1%; *P* = 0.001). Differences in asymptomatic infection rate by age and sex are shown in Table 2.

The highest prevalence of asymptomatic infection was found in the selected villages in Bura (25.0%) and the lowest prevalence was found in villages in Agita (1.5%).

A detailed description of the results obtained by the three methods used by sex, age, and location is shown in Table 3.

### DISCUSSION

Our study shows the presence of asymptomatic *Leishmania* infection among children 4–15 years of age in the districts of Libo Kemkem and Fogera, in the new VL focus of Amhara State in northwestern Ethiopia. The observed overall asymptomatic infection rate was 10.1% (61 of 605). This rate was determined by using a combination of serologic methods, which detected 38 (6.3%) of 605 seropositive persons and LST, which detected 33 (5.6%) of 589 seropositive persons. As proposed initially, the combination of serologic analysis and LST enabled wider detection of asymptomatic infections.

The discordances observed between serologic analysis and LST were expected. Both LST and serologic analysis have been used to assess exposure to *Leishmania*, irrespective of disease presentation, and are frequently used in epidemiologic studies in *Leishmania*-endemic areas. Nevertheless, comparison of LST positivity and seroprevalence rates is complicated by different types of immune response detected by each test. The LST measures a delayed-type hypersensitivity reaction to *Leishmania* and relies on an in vivo cellular immune response to *Leishmania* antigens. Seropositivity is the result of a significant level of *Leishmania*-specific antibodies in the peripheral blood, which is based on a humoral immune response. The two differentiated groups of LST-positive/seronegative and LST-negative/seropositive children observed in our study are consistent with the lack of an association between LST and serologic results. The LST positivity appears later after infection and seems to be a sign of protective immunity against VL. Seropositivity is considered an

### Table 1

<table>
<thead>
<tr>
<th>LST</th>
<th>rK39-ICT†</th>
<th>DAT, no. positive/no. tested (%)</th>
<th>rK39-ICT†</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>+</td>
<td>2/589 (0.3)</td>
<td>6/589 (1.0)</td>
<td>2/589 (0.3)</td>
</tr>
<tr>
<td>-</td>
<td>0/589 (0.0)</td>
<td>23/589 (3.9)</td>
<td>4/589 (0.7)</td>
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<tr>
<td></td>
<td>31/589 (5.3)</td>
<td>558/589 (95.3)</td>
<td>Total = 589 (100)</td>
</tr>
</tbody>
</table>

* rK39-ICT = rK39-immunochromatographic test; DAT = direct agglutination test; LST = leishmanin skin test.
† Global rK39-ICT positive = 8 (1.4%).
Asymptomatic infection rates for visceral leishmaniasis detected by three tests and their combination by sex, age, and location, Ethiopia*  

<table>
<thead>
<tr>
<th>Variable</th>
<th>rK39-ICT, no. (%)</th>
<th>DAT, no. (%)</th>
<th>LST,† no. (%)</th>
<th>Seropositive, no. (%)</th>
<th>Infected, no. (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Whole population (n = 605)</td>
<td>9 (1.5)</td>
<td>32 (5.3)</td>
<td>33 (5.6)</td>
<td>38 (6.3)</td>
<td>61 (10.1)</td>
</tr>
<tr>
<td>Sex</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Boys (n = 309)</td>
<td>7 (2.3)</td>
<td>22 (7.1)</td>
<td>23 (7.6)</td>
<td>28 (9.1)</td>
<td>43 (13.9)</td>
</tr>
<tr>
<td>Girls (n = 296)</td>
<td>2 (0.7)</td>
<td>10 (3.4)</td>
<td>10 (3.5)</td>
<td>10 (3.4)</td>
<td>18 (6.1)</td>
</tr>
<tr>
<td>Age group (years)</td>
<td></td>
<td></td>
<td></td>
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<td></td>
</tr>
<tr>
<td>&lt;5 (n = 77)</td>
<td>1 (1.3)</td>
<td>2 (2.6)</td>
<td>1 (1.3)</td>
<td>2 (2.6)</td>
<td>2 (2.6)</td>
</tr>
<tr>
<td>5–9 (n = 295)</td>
<td>3 (1.0)</td>
<td>13 (4.4)</td>
<td>8 (2.8)</td>
<td>15 (5.1)</td>
<td>21 (7.1)</td>
</tr>
<tr>
<td>10–15 (n = 233)</td>
<td>5 (2.1)</td>
<td>17 (7.3)</td>
<td>24 (10.6)</td>
<td>21 (9.0)</td>
<td>38 (16.3)</td>
</tr>
<tr>
<td>Sub-district</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Agita (n = 133)</td>
<td>0 (0.0)</td>
<td>2 (1.5)</td>
<td>0 (0.0)</td>
<td>2 (1.5)</td>
<td>2 (1.5)</td>
</tr>
<tr>
<td>Bura (n = 140)</td>
<td>3 (2.1)</td>
<td>17 (12.1)</td>
<td>23 (16.5)</td>
<td>18 (12.9)</td>
<td>35 (25.0)</td>
</tr>
<tr>
<td>Dibasifatra (n = 133)</td>
<td>5 (3.8)</td>
<td>3 (2.2)</td>
<td>4 (3.1)</td>
<td>7 (5.3)</td>
<td>10 (7.5)</td>
</tr>
<tr>
<td>Rib Gebriel (n = 132)</td>
<td>1 (0.7)</td>
<td>7 (5.3)</td>
<td>6 (4.6)</td>
<td>8 (6.1)</td>
<td>11 (8.3)</td>
</tr>
<tr>
<td>Yifag (n = 67)</td>
<td>0 (0.0)</td>
<td>3 (4.5)</td>
<td>0 (0.0)</td>
<td>3 (4.5)</td>
<td>3 (4.5)</td>
</tr>
</tbody>
</table>

* rK39-ICT = rK39-immunochromatographic test; DAT = direct agglutination test; LST = leishmanin skin test.
†For 589 children.
of associations between malnutrition, *Leishmania* infection, and active disease. These tools have been locally evaluated and shown to be applicable under field conditions. They will enable investigators to conduct epidemiologic surveillance, thus contributing to ongoing efforts of the Amhara State Health Bureau to control spread of VL to adjacent areas.

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**REFERENCES**


