Almost half a century ago it was recognized that most people living in developing countries had an abnormality of the small intestine (Figure 1). Histologically, this lesion was characterized by flattened villi and an inflammatory infiltrate, much like a milder form of celiac disease, but the vast majority of people appeared healthy. The disorder was labeled “tropical enteropathy” (TE) and was hypothesized to arise from unhygienic environmental conditions, because among migrants relocating to developed nations the condition slowly resolved. Some 30 years after these original observations, a series of studies in different populations around the world showed that TE was present across the tropics, but was absent in some tropical populations of high socio-economic status such as Qatar and Singapore. These studies used thin-layer chromatography to measure fractional excretion of four non-metabolizable sugars (xylose, rhamnose, lactulose, and 3-O-methyl D-glucose) as an assessment of intestinal permeability, although several groups assessed permeability using only lactulose and mannitol, which can be measured at higher throughput. Using this simpler approach, TE was revisited in studies from the Gambia that set out to explore the relationship between enteropathy and poor growth in infancy. From as early as 3–6 months of age, infants had increased intestinal permeability (raised L/M ratio), which correlated inversely with weight and length growth. This work refocused interest on TE, from being an incidental observation among people living in conditions of poverty, to being a disorder potentially underlyng poor growth, which is so frequently observed among children in developing countries. However, direct demonstration of causality was not possible, nor was it established by what mechanism increased intestinal permeability might lead to growth failure.

THE IMPORTANCE OF THE INTESTINAL BARRIER

The intestinal mucosa has to maintain a barrier between the body’s largest microbial ecosystem (containing ~100 trillion organisms) and the sterile bloodstream. Intestinal barrier function comprises several distinct components: first, a mechanical barrier formed by the single layer of epithelial cells, which possess a continuous brush border and are joined by adherens junctions and tight junctions; second, an antimicrobial barrier, composed of defensins, immunoglobulins, and mucins secreted by the epithelial cells; third, an immunological barrier due to innate pattern recognition receptors in the mucosa together with immune cells in the subepithelial layer; and fourth, an ecological barrier created by the gut microbiota that deters pathogens. Disruption of this barrier increases intestinal permeability, enabling luminal contents, including bacteria and microbial-associated products, to leak into the systemic circulation, a process termed microbial translocation (MT). Probes used as markers of abnormal permeation, such as lactulose, have an intermediate molecular size (molecular weight 150–400 Da) and clearly do not reflect the pore size required for translocation of lipopolysaccharide (LPS) (>10,000 Da) or a whole bacterium (typical size 0.5 × 2 μm), but it is believed that when lactulose permeation is increased, a “leaky pathway” is established that enables macromolecules to cross the intestinal barrier.

In the Gambia, researchers went on to show that infants had evidence of endotoxin (LPS) in the blood, together with anti-endotoxin core antibodies (EndoCAb), which correlated with the degree of intestinal permeability. Furthermore, there was other evidence of chronic, low level immunostimulation, including raised lymphocyte and platelet counts, elevated C-reactive protein, and high plasma immunoglobulins, which correlated inversely with growth. Thus, a biologic model emerged in which infants living in conditions of poor sanitation and hygiene develop a chronic inflammatory enteropathy soon after birth, leading to reduced intestinal barrier function and passage of microbial-associated products across a “leaky gut” into the systemic circulation, where they chronically activate the immune system, leading to impaired growth.

WHAT CAUSES TROPICAL ENTEROPATHY?

Tropical enteropathy is a misnomer; although intestinal pathology varies geographically, TE is not confined to the tropics, and not all people in the tropics have enteropathy (see above). It is environmental conditions, rather than the
had a marginal impact,15 leading researchers to consider other potential pathways leading to stunting.16

Between 1978 and 1998 in Africa, Asia, and the Americas, showed a cumulative impact of diarrheal episodes on stunting by 2 years of age, with a similar effect across all studies.21 Interventions to improve hand washing, sanitation, and hygiene have been shown to reduce episodes of diarrhea.22 However, the estimated impact of improved sanitation and hygiene, achieved at 99% coverage, is only a 2.4% reduction in prevalence of stunting when modeled entirely through reductions in diarrhea.23 This modest impact may be because children show “catch up” growth between acute diarrheal episodes24,25; in contrast, children living in poor conditions do not recover from the chronic effects of EE, which may therefore be a more important pathway to stunting than is diarrhea.5

Anemia is a similar prevalent and intractable problem of young children in developing countries that often co-exists with stunting and whose etiology is frequently unexplained.26 The same chronic inflammatory pathway that is hypothesized to link enteropathy and stunting may plausibly lead to an anemia of chronic disease (Figure 2), although this potential mechanism has received little attention to date.

Environmental enteropathy may also reduce the efficacy of oral vaccines, with major public health implications for children living in countries where efficacious vaccines are most needed.27 Although oral rotavirus vaccination is now recommended by the World Health Organization (WHO), its efficacy against severe rotavirus gastroenteritis in multicountry trials was far lower in sub-Saharan Africa (39.3%, 95% confidence interval [CI] = 19.1–54.7)28 and Asia (48.3%, 95% CI = 22.3–66.1)29 than in Europe/USA, where overall efficacy is 85–98%.30,31 The impact of EE on reducing vaccine efficacy may occur either through induction of regulatory T cells, which dampen the vaccine-specific immune response, or through destruction of the live attenuated vaccine by an over-vigorous local immune response.27 Alternatively, EE and reduced vaccine efficacy may both be effects of an extensive immunological disturbance. Data from a birth cohort in Dhaka, Bangladesh, in which 42% of children were stunted by 12 months of age, illustrate the link between undernutrition, enteropathy, and poor vaccine responses. Children who were stunted (HAZ < –2) by 6 months of age had significantly lower oral polio vaccine (OPV) responses at 1 year of age, compared with those who were not stunted by 6 months of age. Furthermore, there was a negative correlation between OPV serotype 3 responses and serum EndoCAb levels, suggesting that both enteropathy and undernutrition may be associated with OPV failure.32 The ongoing PROVIDE trial (www.clinicaltrials.gov identifier NCT01375647) is exploring the biologic basis for underperformance of enteric vaccines among children in Bangladesh.

Figure 1. Endoscopic appearance of environmental enteropathy. Endoscopic view of second part of the duodenum showing villi with characteristic changes of environmental enteropathy: fusion of villi so that instead of a finger-like appearance they take on a leaf-shaped appearance. Sometimes villous fusion goes further and takes on a cerebriform (sulcus and gyrus) appearance. Endoscopic image from Endoscopy Unit, University Teaching Hospital, Lusaka (P. Kelly).

THE IMPACT OF ENTEROPATHIES ON CHILDREN IN THE DEVELOPING WORLD

Stunting (low height-for-age [HAZ]) affects 32% of children living in developing countries and has a major impact on child health and development.7 Chronic undernutrition contributes to one-third of mortality in children < 5 years of age, leads to fewer years at school, reduced adult productivity, and an increased risk of stunting in subsequent generations.7 The first 2 years are a critical time for linear growth and, in particular, for brain growth and development of complex neural networks. Stunting develops between ~9 months (fetal life) and +24 months (termed the critical first thousand days), and its effects are irreversible. Nutritional interventions have only had a marginal impact,15 leading researchers to consider other potential pathways leading to stunting.

Diarrhea is frequent among children living in conditions of poor sanitation and hygiene and has been shown to impact growth.16–20 A pooled analysis of nine studies, conducted
and function. More recently, attention has refocused on the impact of HIV on the gut, following the observation that damage to the intestine may modulate HIV disease progression even before symptomatic gastrointestinal disease.

HIV enteropathy is indistinguishable qualitatively from EE, and where these two enteropathies co-exist they differ only in degree. In a large quantitative analysis of 238 asymptomatic adults biopsied repeatedly over 3 years, crypt depth but not villous height was influenced by HIV; changes in absorption and permeability were only seen in late-stage HIV infection (CD4 count < 200 cells/μL). In patients with HIV-related diarrheal disease, changes in villous height were much more pronounced, irrespective of the infectious diagnosis; villous height correlated positively with nutritional status and negatively with markers of circulating TNF pathway activation. To our knowledge, eight published studies have reported morphometric (N = 6) and/or permeability (N = 4) data in both early and late HIV infection. Of the morphometric studies, four found no evidence of villous changes in early HIV infection but two did. Crypt changes were observed in early infection in two but not in others; generally it was the larger studies that found the difference. Importantly, of the permeability studies, none found any evidence of enteropathy in early HIV. Taken together, although there are some enteropathic changes (crypt hyperplasia, cytokine dysregulation) in early HIV infection, the most important changes from the perspective of MT are predominantly seen in advanced disease.

It is well recognized that mucosal T cell depletion occurs in HIV infection. Recent studies have defined this immunological hit in more detail: the Th17 population of CD4 cells is most severely depleted and this occurs earlier in HIV infection than might have been suspected before refined techniques to define T cell subsets became available. Because Th17 cells play a major role in defense against bacterial, fungal, and parasitic infections, this may explain the spectrum of intestinal pathology seen in HIV infection. There is evidence that HIV itself can directly disrupt mucosal integrity of primary epithelial cells in vitro by down-regulating tight junction proteins, leading to increased intestinal permeability. Brenchley and others showed that mucosal damage enables MT to occur in HIV-infected individuals. In their study, the level of circulating microbial products correlated with the degree of immune activation, which is the hallmark of HIV infection and best indicator of disease progression. Intriguingly, sooty mangabeys, who develop non-pathogenic simian immunodeficiency virus infection, preserved Th17 cells in the gut and did not show signs of MT or immune activation. However, it remains debatable whether MT is a cause or consequence of advanced HIV disease. Several studies

![Diagram of proposed causal pathway linking enteropathy with adverse health outcomes in developing countries.](image-url)
have confirmed the finding of higher levels of LPS in HIV-infected, compared with HIV-uninfected, individuals, and that LPS levels do not completely normalize despite effective antiretroviral therapy (ART) \textsuperscript{46–48}; however, in these studies the level of MT did not correlate with HIV disease progression, whether assessed by viral load/CD4 count, \textsuperscript{46} proportion of activated T cells, \textsuperscript{48} or speed of clinical disease progression. \textsuperscript{47} In contrast, a recent case-control study of HIV-infected individuals in the SMART trial showed that those with the highest quartile of soluble CD14 levels, a marker of monocyte responsiveness to LPS, had a 6-fold higher risk of death compared with those in the lowest quartile. \textsuperscript{48} Taken together, although there is undoubtedly evidence of increased MT in the context of HIV infection, its contribution to disease progression remains uncertain. HIV enteropathy may be a modulator of disease progression, which is primarily driven by viral replication. Virologic suppression with ART is the most effective treatment to reduce the immune activation that underlies disease progression. However, residual MT and immune activation frequently persist, despite virologic control, and are associated with increased morbidity and a poor immunologic response to ART. \textsuperscript{50}–\textsuperscript{52} Current trials (reviewed below) are exploring whether adjunctive interventions to improve enteropathy may modulate HIV disease outcome in the era of ART.

THE OVERLAPPING AND INTERACTING NATURE OF ENTEROPATHIES

The HIV epidemic affects populations already blighted by chronic stunting and food insecurity. HIV and malnutrition overlap, and the interacting effects of infection, enteropathy and immunodeficiency propagate morbidity and mortality. The spectrum of enteropathies caused by HIV, poor sanitation/hygiene, persistent diarrhea, inadequate diet, and severe acute malnutrition (SAM) compound each other (Figure 3). The overlapping and interacting nature of enteropathies was elegantly demonstrated in an early study from the Gambia, \textsuperscript{53} in which intestinal permeability was shown to rise progressively in a comparison of asymptomatic children (mean L/M ratio 0.42), well but undernourished children (L/M 0.52), malnourished children (L/M 1.3), and children with persistent diarrhea (L/M 2.85). Although the children who showed clinical recovery demonstrated improvement in intestinal integrity, many children do not recover from the persistent diarrhea-malnutrition cycle, particularly in the context of HIV infection where immunodeficiency and HIV enteropathy compound the situation further.

Malnutrition has therefore been viewed as an enteric infectious disease because of the vicious cycle in which diarrhea exacerbates malnutrition and malnutrition exacerbates diarrhea (reviewed in Reference 54). The impact of diarrheal episodes on weight and height has been clearly documented in multiple field studies. In a Guatemalan study undertaken in the 1960s, Mata \textsuperscript{55} elegantly demonstrated the progressive decline across growth centiles that occurred with each successive episode of diarrhea during early childhood. Most recently, the Global Enterics Multi-Center Study has provided new data on the profound sequelae of diarrheal episodes among children 0–59 months of age in seven African/Asian countries. Compared with age, gender, and location-matched controls, cases had a significant decrease in linear

![Figure 3](image-url). Overlapping causes of enteropathy in the developing world. There are multiple underlying causes of enteropathy among people living in developing countries, which overlap on a background of environmental enteropathy. Environmental enteropathy is almost universal among people living in conditions of poverty. Overlapping causes of enteropathy shown here are based on known associations, but the size of each overlap is speculative and is not meant to represent prevalence. MN = micronutrient; SAM = severe acute malnutrition.
growth and a striking 7-fold (rate ratio 7.1; 95% CI 5.0, 10.1) increase in mortality 60 days after an episode of moderate-severe diarrhea. It is similarly well recognized that malnutrition impacts diarrhea. In an intensive prospective study of 61 children < 5 years of age in a Brazilian urban slum, children with malnutrition, measured either as low weight-for-age or low HAZ, had an increased incidence and duration of diarrhea in the subsequent 2 months, compared with better nourished children. The overlapping impact of enteric infection and malnutrition likely arises from a combination of impaired intestinal barrier function together with reduced capacity for mucosal repair caused by poor nutrient stores; reduced absorptive capacity, and nutrient loss, particularly in cases of persistent diarrhea; and reduced appetite in the face of increased catabolic demands.

The reciprocal interactions between malnutrition and enteropathy are well illustrated in a recent Dhaka birth cohort study. Enteropathy was shown to influence malnutrition, because both prolonged diarrhea during infancy and impaired intestinal barrier function (as measured by elevated serum EndoCAb at 6 months of age) were associated with stunting at 1 year of age; conversely, malnutrition was shown to influence enteropathy, because children who were born stunted were more likely to develop severe diarrhea, infection with Entamoeba histolytica, and diarrhea caused by Cryptosporidium and enterotoxigenic Escherichia coli during the first year of life. This study highlights the importance of preventing or treating enteropathy to reduce malnutrition, and the inter-generational impact of maternal undernutrition on infant enteropathy and malnutrition.

OTHER ENTEROPATHIES IN THE DEVELOPING WORLD

Dietary factors also exacerbate enteropathy among children living in developing countries. Zinc deficiency, estimated to affect more than 20% of people worldwide, causes structural intestinal changes characterized by villous atrophy and reduced crypt cell production rate. Zinc deficient children experience increased diarrheal morbidity, supplementation with zinc reduces the severity and duration of both acute and persistent diarrhea. EE may also exacerbate zinc deficiency by reducing resorption of endogenous zinc from the small intestine, which is critical for zinc homeostasis. Thus, zinc deficiency and EE are interacting factors that may propagate overt clinical disease in children with overlapping causes of enteropathy. Children with poor quality diets may also be exposed to aflatoxin, a fungal metabolite that contaminates inadequately stored crops such as maize and peanuts. Aflatoxin impairs intestinal integrity in animal models and is associated with stunting in children, potentially through the same pathway as EE.

It is also apparent that some emerging diseases of the developing world are associated with an underlying enteropathy. Obesity, diabetes, and the metabolic syndrome are all characterized by increased intestinal permeability. Mice receiving a high-fat diet have raised LPS levels, and the resulting immune activation causes insulin resistance through cytokine-mediated phosphorylation of insulin receptor. Given the rising prevalence of these “developed” diseases in areas of the developing world that are undergoing economic transition, enteropathies become an even more important pathological process in this setting that require further recognition and research.

DIAGNOSIS, PREVENTION, AND TREATMENT OF ENTEROPATHIES

Several large programs are currently addressing priority research and implementation questions related to intestinal infections and EE. The MAL-ED project (http://mal-ed.fnih.org/) is investigating the link between EE and malnutrition among children in eight countries. A panel of investigations conducted on longitudinal birth cohorts recruited at each site are assessing intestinal inflammation and barrier function, enteric infections, growth/development, and vaccine responses, to better inform rational design of intervention strategies to break the enteropathy-malnutrition cycle. One of the major challenges in young children, in the absence of intestinal biopsy material, is the paucity of well-validated, non-invasive biomarkers to assess enteropathy. The MAL-ED program is evaluating a range of enteropathy markers in urine, feces, and blood that may be associated with poor growth in early childhood.

Several large field programs currently underway are investigating strategies to reduce malnutrition and/or diarrheal disease among children in developing countries. Alive & Thrive (www.aliveandthrive.org) is a 5-year initiative underway in Ethiopia, Bangladesh, and Viet Nam, aimed at improving exclusive breastfeeding and complementary feeding practices in the first 2 years of life. These strategies have proven efficacy in reducing infant mortality, reducing frequency and severity of diarrhea, and improving long-term nutritional status, but are currently poorly adopted in many settings. Two trials due to start recruitment shortly in Zimbabwe (SHINE) and Kenya/Bangladesh (WASH Benefits) will test the hypothesis that improved sanitation/hygiene from birth will prevent or ameliorate EE and reduce childhood stunting. Another cluster-randomized trial underway in Orissa, India, (www.clinicaltrials.gov identifier NCT01214785) is investigating the impact of latrine construction and use on diarrheal disease, intestinal helminth infections, and nutritional status. A further trial in Malawi (NCT01440608) is investigating the effectiveness of high-dose zinc treatment or deworming with albendazole on intestinal permeability in rural 1–3 year old children at high risk of EE.

Previous trials have generally shown disappointing results with regards to treatment of underlying enteropathy in children with malnutrition, although interpretation of trial results is partly hindered by the absence of well-validated markers. Trehan and others conducted a double-blind, placebo-controlled trial of rifaximin, a non-absorbable antibiotic, in 3–5 year old rural Malawian children with EE. After 7 days of rifaximin or placebo, administered under direct observation of study staff, intestinal permeability as measured by the L/M ratio was not significantly different between groups, although no measures of MT were untaken in this study. The same research group undertook a trial of the probiotic Lactobacillus GG in rural Malawi, but again demonstrated no difference in intestinal integrity between groups after 30 days of probiotic or placebo. A probiotic and prebiotic functional food, given to Malawian children with SAM following stabilization, showed no improvement in nutritional cure compared with control. However, this does not necessarily imply that
these approaches are futile. Choice of antibiotic/probiotic, treatment duration, age group, and primary outcome may all impact results, and some of these strategies continue to be explored in future trials.

Several trials in HIV-infected individuals are investigating strategies to target enteropathy and reduce immune activation. One trial (NCT01090102) is using 5-aminosalicylic acid, an oral anti-inflammatory agent widely used in inflammatory bowel disease, to reduce enteric inflammation. Other trials are aiming to reduce MT, using either probiotics (NCT01184456), a prebiotic/probiotic mixture (NCT00688311), or the non-absorbed antibiotic rifaximin (ACTG trial 5286). It is hoped that results of these trials will clarify the role of enteropathy in HIV infection and may inform choice of adjunctive therapies, particularly for HIV-infected individuals with poor immunologic response to ART, in whom residual immune activation is known to be contributory.80

**NEXT STEPS**

There is a fundamental need to better understand the basic science underlying enteropathies. The interaction between enterocytes, pathogens, and the mucosal immune system is likely to be critical in development and resolution of enteropathy. It is also becoming apparent that the intestinal microbiota has a profound impact on gut structure and function and nutritional status. Developments in high-throughput sequencing and gnotobiotic mouse models now enable characterization of the composition and function of gut microbial communities.80 Food type and nutrient load both impact the gut microbiota; conversely, changes in the gut microbiota impact nutrient use and energy balance.80 Transfer by gavage of the microbiota from a child with SAM into a gnotobiotic mouse causes malnutrition in the animal, and therapeutic feeding causes shifts in the composition and functional profile of the gut microbial community, demonstrating the bidirectional interaction between the microbiota and nutritional status.81

In summary, there is a strong rationale for developing interventions that prevent or ameliorate enteropathy, with the aim of reducing stunting, anemia, malnutrition, and improving HIV disease outcome. Such interventions have the potential to improve the health of millions of people in developing countries, especially among children <5 years of age, who are most affected by the overarching epidemics of stunting, malnutrition, and HIV.

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Authors’ addresses: Andrew Prendergast, Centre for Paediatrics, Blizard Institute, Queen Mary University of London, UK, E-mail: a.prendergast@qmul.ac.uk. Paul Kelly, Centre for Digestive Diseases, Blizard Institute, Queen Mary University of London, UK, E-mail: m.p.kelly@qmul.ac.uk.

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