Hepatitis E virus (HEV) is a hepatotropic virus that causes acute self-limiting hepatitis. This virus is the leading cause of outbreaks and sporadic acute hepatitis in industrialized countries. In developed countries, sporadic infections predominate and are usually considered to be imported through travel to disease-endemic countries. The virus is classified into four major genotypes (1, 2, 3, and 4) and has one serotype. All four virus genotypes infect humans. Generally genotypes 1 and 2 cause outbreaks and sporadic, water-borne infections, and genotypes 3 and 4 cause sporadic foodborne infections. Recently, more autochthonous infections are being reported from industrialized countries. This finding may be partially caused by zoonotic HEV infections. The first animal strain of HEV was isolated from pigs in the mid-western United States and its nucleotide sequence has been determined. Swine HEV has since been identified in all swine-producing countries worldwide. HEV sequences identified from swine belong to either genotype 3 or 4.

Hepatitis E virus is considered enzootic worldwide; infected swine appear clinically normal but show microscopic evidence of hepatitis. Studies have shown experimental cross-species infection of primates by swine HEV and of pigs by human HEV. In a study in the United States, increased prevalence of antibodies against HEV was observed in pig handlers compared with controls. High genetic relatedness between human and swine HEV was observed in countries such as United States, Taiwan, Japan, China, and countries in Europe. These studies provide evidence for pigs as reservoirs of HEV infection in humans. However, in India, HEV infecting humans belongs to genotype 1 and swine are infected by genotype 4. Therefore, hepatitis E may not be a zoonotic disease, although high antibody (96.5%) prevalence rates have been demonstrated in pigs. The objective of our study was to detect and characterize HEV from swine and swine handlers in Vellore, southern India and to assess the possibility of human infection caused by swine HEV.

A total of 102 swine serum samples (60 males and 42 females) were obtained from slaughter houses in and near Vellore. Plasma samples were obtained from 34 swine handlers (20 males and 14 females) residing in Vellore who had a median (SD) age of 35 (12.8) years. Viral RNA was extracted by using the Trizol method (Invitrogen, Carlsbad, CA), and a reverse transcription–polymerase chain reaction was carried out by using India swine-specific genotype 4 primers and universal primers, respectively. Amplified products were sequenced, and phylogenetic analysis was performed to genotype HEV. An enzyme-linked immunosorbent assay (ELISA) was performed to detect IgG against HEV by using the Genelabs HEV IgG enzymeimmunoassay (EIA) (MP Bio Ltd., Singapore) and the Wantai HEV IgG EIA (Wantai Biological Pharmacy, Beijing, China) for swine handlers. Seroprevalence of antibodies against HEV in age and geographically matched controls were obtained from a previous study that used the Genelabs assay to compare HEV exposure in age-stratified urban and rural populations in Vellore. From the previous study, 200 samples, 100 each from urban and rural populations (91 males and 109 females) with a median (SD) age of 35 (2.5), were tested by using the Wantai assay to determine seroprevalence of antibodies against HEV. Data analysis was carried out by using SPSS version 16 (SPSS Inc., Chicago, IL). Groups were compared using chi-square test/Fisher exact test for categorical data.

Of 102 swine serum samples tested, 2 samples were positive for HEV RNA. They were characterized as genotype 4 (Genbank accession nos. GQ494002 and GQ494003) and clustered with other swine HEV from India (Figure 1). Of 34 plasma samples from swine handlers tested, none were positive for HEV RNA. However, the Wantai assay showed that 94.1% were positive for IgG against HEV. Prevalence in swine handlers was higher than in rural (59%) and urban (73%) adults, respectively ($P < 0.001$). Seroprevalence rates determined by using the Genelabs HEV IgG EIA showed a similar pattern, but lower prevalence rates (Figure 2). The prevalence of 67.6% in swine handlers using the Genelabs assay was also significantly higher ($P < 0.001$) than in rural (20.5%) and urban (35.5%) adults. Seroprevalence rates for rural and urban adults determined by using the Genelabs assay have been reported. In comparison with controls, the swine handlers had higher exposure (odds ratio = 8.2, 95% confidence interval = 1.91–35.42 and odds ratio = 5.34, 95% confidence interval = 2.55–11.18) when determined by using the Wantai and Genelabs kits, respectively.

Our results partially support the finding that genotype 4 HEV circulates in swine but not in humans in southern India,
high HEV exposure with human strains could provide cross-protection against disease, but increased exposure to swine HEV in handlers could result in a boosting of the antibody response. This hypothesis could also explain the differences in disease susceptibility to swine HEV in developing countries and industrialized countries.

It is also important to note that HEV isolated from swine in this study clustered closely with a previously reported zoonotically transmitted genotype 4 strain isolated from a patient in the United Kingdom (Figure 1), who had traveled in India before onset of the disease. These data provide insight into possible non–feco-oral transmission modes of HEV in India and call for increased surveillance for genotype 4 HEV in humans.

Received August 16, 2010. Accepted for publication January 6, 2011.

Authors’ addresses: Rosario Vivek and Gagandeep Kang, Wellcome Trust Research Laboratory, Department of Gastrointestinal Sciences, Christian Medical College, Vellore 632004, India, E-mails: vivekm@cmcvellore.ac.in and gkang@cmcvellore.ac.in.

REFERENCES


