Symptomatic and Asymptomatic Cryptosporidium Infections in Children in a Semi-Urban Slum Community in Southern India


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Abstract. Cryptosporidium is a leading cause of childhood diarrhea in developing countries. We investigated symptomatic and asymptomatic cryptosporidiosis in 20 children less than two years of age in a semi-urban slum in southern India. All surveillance (conducted every two weeks) and diarrheal samples from 20 children (n = 1,036) with cryptosporidial diarrhea previously identified by stool microscopy were tested by polymerase chain reaction–restriction fragment length polymorphism for species and subgenotype determination. Thirty-five episodes of cryptosporidiosis were identified in 20 children, of which 25 were diarrheal. Fifteen episodes were associated with prolonged oocyst shedding. Multiple episodes of cryptosporidiosis occurred in 40% of the children. Most infections were with C. hominis, subtype Ia. Children with multiple infections had significantly lower weight-for-age and height-for-age Z scores at 24 months but had scores comparable with children with a single episode by 36 months. Multiple symptomatic Cryptosporidium infections associated with prolonged oocyst shedding occur frequently in this disease-endemic area and may contribute to the long-term effects of cryptosporidiosis on physical growth in these children.

INTRODUCTION

Cryptosporidium spp. is a leading cause of diarrhea in children in developing countries where it primarily affects those less than five years of age. Diarrhea caused by these parasites in early childhood has been associated with subsequent cognitive function deficits and growth faltering and stunting, and the risk of stunting increases with the number of episodes per year. A study from Peru found that symptomatic and asymptomatic cryptosporidiosis in children were associated with growth faltering after an infection but recovery was slower in children with symptomatic infection. In India, several studies have reported Cryptosporidium spp. in children with diarrhea (ranging from 1.3% to 18.9%) and in asymptomatic children (0–3%). and one study reported up to 9.8% positivity in asymptomatic children (with 13.1% positivity in symptomatic children).

In a previous study on a well-defined birth cohort of 452 children followed-up for 3 years in a semi-urban slum community in Vellore in southern India, 53 children with diarrhea were identified to have Cryptosporidium by microscopic examination of stool samples (cryptosporidial diarrhea). Most (41 of 53) children had cryptosporidial diarrhea at 2 years of age. The most common species associated with diarrhea was C. hominis, and subgenotyping by polymerase chain reaction–restriction fragment length polymorphism (PCR-RFLP) at the polymorphic Cpgp40/15 locus identified Ia as the most common subgenotype. In another study on children hospitalized with diarrhea, we found that PCR detected more than three times the number of cryptosporidial infections detected by microscopy.

METHODS

Study participants and samples. Twenty of 41 children who had at least one episode of cryptosporidial diarrhea identified by microscopy at ≤ 2 years of age were selected for this longitudinal study by using a random numbers list. All children belonged to a birth cohort residing in the semi-urban slum areas of Vellore in southern India. Recruitment and follow-up of the cohort have been described. Surveillance stool samples collected every two weeks, and diarrheal stool samples collected each time the child had a diarrheal episode were stored at –70°C. All available surveillance and diarrheal stool samples for the 20 children collected from birth to two years of age (n = 1,036 samples, 196 diarrheal samples, and 840 surveillance samples) were examined for Cryptosporidium spp. by PCR using the methods described below.

An episode of diarrhea was defined as at least one day of diarrhea (with ≥ 3 watery stools in a 24-hour period) preceded and followed by ≥ 2 days without diarrhea. An episode of cryptosporidiosis was defined as a period during which stool samples were positive for Cryptosporidium by PCR. The episode was considered to have ended when at least two consecutive samples showed negative results. An episode was classified as symptomatic or associated with diarrhea if at least one sample in that episode was a diarrheal sample. Co-infection was defined as identification of Cryptosporidium and at least one other enteric pathogen in a diarrheal stool sample.

Assessment of diarrhea severity. The severity of the diarrheal episode was assessed by using the Vesikari scoring system, which is commonly used to assess severity of rotavirus diarrhea in children. The 20-point score is determined by the total duration of diarrhea, the maximum number of stools passed in 24 hours, the duration of vomiting (if present), the maximum number of vomiting episodes in 24 hours, fever (in °C), and the degree of dehydration. Although the score is designed for acute watery diarrhea caused by rotavirus, total and component assessment have previously yielded useful correlation with severity in a previously published report of cryptosporidiosis in the cohort, of which the 20 study children formed a subset.

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Baseline sociodemographic data. Baseline sociodemographic data including sex, socioeconomic status, religion, family size, number of siblings, maternal education, education, and occupation of the head of the household were available. In addition, a hygiene assessment carried out at least every six months included information elicited by enquiry on hand-washing before feeding the child and after defecation, and by observation on covering drinking water, use of dedicated dippers to obtain drinking water stored in large containers, and washing of vegetables, fruit, and cooking vessels.

Anthropometric measurements. All children had birth weights and lengths recorded, and heights and weights were measured each month for the 36 months of follow-up. Weight-for-age (WAZ) and height-for-age (HAZ) z-scores were calculated by using the 2006 World Health Organization child growth standards as the reference population.

DNA extraction and PCR. DNA extraction using a QIAamp stool DNA extraction kit (Qiagen, Valencia, CA) and nested PCR of the Cryptosporidium small subunit ribosomal RNA (SSU rRNA) locus were performed on all diarrheal and surveillance stool samples. Initially, three sequentially collected surveillance samples from each child were pooled and resulted in 343 pools derived from 1,036 samples, DNA was extracted from the pool and analyzed by PCR. From all positive pools, DNA was re-extracted from individual stool samples, and the infecting species and subgenotype were identified by PCR-RFLP at the SSU rRNA locus and the Cpgp40/15 locus, respectively, by using previously described protocols.

Diarrheal stool samples were also screened for parasite ova and cysts by microscopy, for bacterial pathogens by culture, and for rotavirus by enzyme-linked immunosorbent assay (Rota IDEIA; Dako, Ely, United Kingdom). A subset were also screened for Campylobacter, diarrheagenic Escherichia coli, norovirus genogroups I and II, sapovirus, and adenovirus by PCR.

Statistical analysis. Comparison of diarrheal severity and duration among children with single and multiple episodes of cryptosporidiosis was performed by using the Mann-Whitney U test. Nutritional status among children with single and multiple cryptosporidial infections were compared at the time of weaning (median age = 3 months, interquartile range [IQR] = 1.9–4 months), 24 months, and 36 months of age by using the Mann-Whitney U test. Baseline sociodemographic parameters, hygiene, and breastfeeding history were compared by using Fisher’s exact test for categorical variables and two-tailed t-test or the Mann-Whitney U test for continuous variables.

The study was reviewed and approved by the Institutional Review Board of Christian Medical College, Vellore, and informed consent was obtained from the parents.

RESULTS

Episodes of cryptosporidiosis. A mean (SD) of 42 (9.6) surveillance stool samples and 10 (5.4) diarrheal stool samples per child were examined, and 1,036 samples were tested in 343 pools of 3 consecutive samples each by SSU rRNA PCR. A total of 28 (8.2%) pools were positive, of which 5 (17.9%) were consecutive pools. Twelve (60%) children had more than one positive pool. When the individual stool samples in the positive pools were tested, 87 (8.4%) of 1,036 samples (both diarrheal and asymptomatic) were positive (Figure 1).
Thirty-five episodes of cryptosporidiosis were identified among the 20 children. Two children (child 18 and child 20) had a gap of more than one month between two available positive samples. Intervening samples were not collected because the children were out of the study area at that time, and these were considered to belong to a single episode. One child dropped out of the study at 10 months (child 19). Twenty-two (71.4%) of 30 episodes were associated with diarrhea and the remaining 10 (28.6%) were asymptomatic. One episode of asymptomatic cryptosporidiosis was preceded by symptomatic cryptosporidiosis less than a month earlier (child 6, Table 1).

Most (31 of 35) symptomatic and asymptomatic infections were caused by *C. hominis*. There was one episode each caused by *C. parvum* and *C. meleagridis*. In two episodes (second episode in child 6 and the first of four episodes in child 14), only a faint band was obtained in the SSU rRNA PCR, and the species could not be identified by RFLP analysis. Twenty-two (71%) of 31 *C. hominis* could be subgenotyped by PCR-RFLP at the *Cpgp40/15* locus, and most (12 of 22, 54.5%) were subgenotype Ia as previously documented in this birth cohort (child 6, Table 1).

Thirteen episodes of cryptosporidial diarrhea in 12 children were followed by asymptomatic oocyst shedding for 7–65 days. Six episodes of cryptosporidial diarrhea in six children were preceded by asymptomatic oocyst shedding that ranged from 7 to 22 days before the onset of diarrhea (Figure 1). In four of six children with asymptomatic oocyst shedding before diarrhea, the diarrheal stool samples were positive for *Cryptosporidium* and other enteric pathogens, including rotavirus (child 4), enterotoxigenic *E. coli* (child 16), and adenovirus (child 13 and child 19). Two children with intermittent diarrhea during a cryptosporidial episode were found to be co-infected with adenovirus and *Giardia* (child 5) and enteroaggregative *E. coli* (child 6).

**Multiple episodes.** Multiple episodes of cryptosporidiosis infection occurred in 40% (8 of 20) of the children less than two years of age; three children were infected three times and two children had four episodes each (Table 1). In all the eight children with multiple infections, the first infection was symptomatic, and three of eight second infections, one of five third infections, and zero of two fourth infections were associated with diarrhea. In all but two children with multiple episodes of cryptosporidiosis, subsequent episodes were asymptomatic or associated with a decrease in duration and severity of diarrhea (Table 1). Both children with greater severity during the second episode of cryptosporidial diarrhea were co-infected with other enteric pathogens (adenovirus and enterotoxigenic *E. coli* in child 14 and adenovirus and rotavirus in child 20).

Almost all recurrent episodes of cryptosporidiosis occurred more than a month after the previous episode (median = 91 days, IQR = 22–462 days). The median duration between a diarrheal episode and an asymptomatic episode of cryptosporidiosis was 242.5 days (IQR = 42–462 days).

In children with multiple episodes of cryptosporidiosis, almost all (13 of 15) episodes after the first one were caused by *C. hominis*, and only one child (child 20) had *C. meleagridis* infection after three *C. hominis* infections (Table 1). Subgenotypes could be determined for 6 of 8 primary infections but only for 4 of 15 later infections. Thus, subgenotypes were identified for initial and subsequent infections in only three of eight children with multiple infections. Two children (child 8 and child 18) were re-infected with the same subtype (Id and Ia, respectively), and the third child (child 11) had an initial subtype Ib infection and was re-infected with subtype Ia.

When clinical, sociodemographic, and environmental parameters were compared between children with single and multiple episodes of cryptosporidiosis (Table 2), children with multiple

### Table 1

<table>
<thead>
<tr>
<th>Child no.</th>
<th>First episode</th>
<th>Species, subtype</th>
<th>Interval (days)</th>
<th>Second episode</th>
<th>Species, subtype</th>
<th>Interval (days)</th>
<th>Third episode</th>
<th>Species, subtype</th>
<th>Interval (days)</th>
<th>Fourth episode</th>
<th>Species, subtype</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>S</td>
<td>Ch, Ie</td>
<td>42</td>
<td>A</td>
<td>Ch, ND</td>
<td>28</td>
<td>A</td>
<td>Ch, ND</td>
<td></td>
<td></td>
<td>Ch, ND</td>
</tr>
<tr>
<td>6</td>
<td>S</td>
<td>Ch, Ia</td>
<td>167</td>
<td>A</td>
<td>ND</td>
<td>8</td>
<td>A</td>
<td>Ch, ND</td>
<td></td>
<td></td>
<td>Ch, ND</td>
</tr>
<tr>
<td>8</td>
<td>S</td>
<td>Ch, Id</td>
<td>34</td>
<td>S</td>
<td>Ch, Id</td>
<td>88</td>
<td>A</td>
<td>Ch, ND</td>
<td></td>
<td></td>
<td>Ch, ND</td>
</tr>
<tr>
<td>11</td>
<td>S</td>
<td>Ch, Ib</td>
<td>462</td>
<td>A</td>
<td>Ch, Ia</td>
<td>204</td>
<td>A</td>
<td>Ch, ND</td>
<td></td>
<td></td>
<td>Ch, ND</td>
</tr>
<tr>
<td>13</td>
<td>S</td>
<td>Ch, Ia</td>
<td>318</td>
<td>A</td>
<td>Ch, ND</td>
<td>202</td>
<td>S</td>
<td>Ch, ND</td>
<td>49</td>
<td>A</td>
<td>Ch, ND</td>
</tr>
<tr>
<td>14</td>
<td>S</td>
<td>ND</td>
<td>85</td>
<td>S</td>
<td>Ch, Ie</td>
<td>383</td>
<td>A</td>
<td>Ch, ND</td>
<td>22</td>
<td>A</td>
<td>Ch, ND</td>
</tr>
<tr>
<td>18</td>
<td>S</td>
<td>Ch, Ia</td>
<td>394</td>
<td>A</td>
<td>Ch, Ia</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>20</td>
<td>S</td>
<td>Ch, ND</td>
<td>91</td>
<td>S</td>
<td>Ch, ND</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

* S = symptomatic (diarrhea); Ch = Cryptosporidium hominis; A = asymptomatic; ND = not determined; Cm = C. meleagridis.

### Table 2

<table>
<thead>
<tr>
<th>Variable</th>
<th>Single infection (n = 12)</th>
<th>Multiple infections (n = 8)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Male sex</td>
<td>4</td>
<td>4</td>
<td>0.648*</td>
</tr>
<tr>
<td>Low socioeconomic status</td>
<td>7</td>
<td>4</td>
<td>1.000*</td>
</tr>
<tr>
<td>No formal education for head of family</td>
<td>3</td>
<td>3</td>
<td>0.642*</td>
</tr>
<tr>
<td>No formal education for mother</td>
<td>4</td>
<td>2</td>
<td>1.000*</td>
</tr>
<tr>
<td>Mean (SD) maternal age at birth (years)</td>
<td>22.75 (4.33)</td>
<td>24.88 (2.42)</td>
<td>0.225†</td>
</tr>
<tr>
<td>Mean (SD) family size</td>
<td>6.08 (1.62)</td>
<td>6.25 (1.28)</td>
<td>0.810</td>
</tr>
<tr>
<td>Mean (SD) no. of siblings</td>
<td>2.1 (1.28)</td>
<td>3 (0.89)</td>
<td>0.094‡</td>
</tr>
<tr>
<td>Mean (SD) birth weight (kg)§</td>
<td>2.97 (0.49)</td>
<td>2.96 (0.24)</td>
<td>0.931‡</td>
</tr>
<tr>
<td>Mean (SD) duration of exclusive breastfeeding (months)</td>
<td>1.59 (0.82)</td>
<td>3.89 (1.26)</td>
<td>0.001‡</td>
</tr>
<tr>
<td>Mean (SD) no. of diarrheal episodes</td>
<td>5.42 (3.61)</td>
<td>7.5 (3.16)</td>
<td>0.173§</td>
</tr>
<tr>
<td>Animals in the household</td>
<td>1</td>
<td>2</td>
<td>0.537*</td>
</tr>
<tr>
<td>Covered drinking water container</td>
<td>9</td>
<td>4</td>
<td>0.356*</td>
</tr>
<tr>
<td>Mother washed hands before feeding the child</td>
<td>4</td>
<td>5</td>
<td>0.362*</td>
</tr>
</tbody>
</table>

* By Fisher’s exact test.
† By two-tailed t-test.
‡ By Mann-Whitney U test.
§ Data missing for one child.
episodes were exclusively breastfed for a longer duration than children with a single episode ($P < 0.001$).

When the 11 children with a single episode (1 of the 12 dropped out of the study at 10 months of age) and 8 children with multiple episodes of cryptosporidiosis were compared, there were no significant difference in birth weight or in WAZ and HAZ scores at the time of weaning (median = 2 months, IQR = 1–3 months). However, by 24 months of age, children with multiple episodes of cryptosporidiosis had significantly lower $Z$ scores for WAZ and HAZ than those with single infections ($P = 0.013$ and $P = 0.021$, respectively) (Figure 2).

By 36 months of age, there were no significant differences in WAZ or HAZ scores between children with single and multiple infections, which indicated recovery from malnutrition.

**DISCUSSION**

Using molecular tools and a longitudinal study design, we found high rates of multiple cryptosporidial infections, prolonged oocyst shedding before and after an episode of cryptosporidiosis, and a greater association of growth faltering with multiple infections in children in this semi-urban slum community in southern India. Other studies of children in developing countries have reported multiple infections in 14–30% of cases. Longitudinal studies from Brazil and Israel and cross-sectional studies on children with diarrhea from Uganda and Mexico have reported a greater prevalence of asymptomatic infections than asymptomatic infections. In contrast, a longitudinal study from Peru consistently identified more asymptomatic infections than symptomatic infections. Both longitudinal studies from Brazil and Israel and those from Guinea-Bissau and Uganda identified more persistent (diarrhea lasting ≥ 14 days) diarrhea than acute diarrhea (diarrhea lasting less than 4 days). However, no persistent diarrhea was found in this study.

Occurrence of multiple infections, including repeated episodes of diarrhea, could be caused by infection with different species or subtypes. However, this finding was not supported by our study, where all re-infections, except one, were with *C. hominis*, and further characterization identified the same subgenotype in two of three children in whom all subgenotyping data were available. In Peru, *C. hominis* was detected in 6 of 15 children with 2 episodes and 1 of 2 children with 3 infections, and other re-infections were initially with a potentially zoonotic species followed by *C. hominis*. The studies are not directly comparable because of differences in study design and typing methods, but it is interesting to speculate on geographic differences in exposure to different cryptosporidial species and host response to infection.

This study did not evaluate any immunologic parameter. However, the data supports a decrease in frequency of infections and symptoms with age, probably caused by development of immunity. It would have been interesting to have more data on the subgenotypes infecting children with multiple episodes to elucidate whether protection is species or subgenotype specific. We were able to subgenotype 75% of primary infections but only 23% of post-primary infections (Table 1). These data may be partially explained by the fact that most of these were asymptomatic infections in which parasite burden is likely to be lower. The PCR-RFLP for the SSU rRNA gene (which is a multicopy gene) that is used for species determination is more sensitive than the PCR-RFLP for the *gp40/15* gene (which is a single-copy gene) used for subgenotyping. This finding may explain why it was possible to determine species but not subgenotype in these infections. In two cases, species could not be identified by the SSU rRNA PCR, possibly because of the low oocyst burden.

Our study documented prolonged asymptomatic oocyst shedding before and after cryptosporidial diarrhea in at least 50% of children studied. These findings may have important implications for transmission of disease and long-term effects on growth. The finding of significantly greater growth faltering at 24 months of age in children with multiple infections must be interpreted with caution given the small number of children included in the study. However, this finding is biologically plausible and supports previous studies that have shown deleterious effects on growth and development after symptomatic and asymptomatic infections in children who acquired the infection at less than one year of age and with boys rather than girls affected more significantly. In a longitudinal study from Brazil, *C. hominis* and *C. parvum* infections were associated with decrease in HAZ scores within three months post-infection but this decrease was found to persist at 3–6 months after *C. hominis* infections, but not *C. parvum* infections.

Low birth weight, malnutrition, stunting and lack of breastfeeding have been reported to predispose children to cryptosporidiosis. In this birth cohort, the median age of weaning was three months (IQR = 1.8–4 months). In our study, multiple infections were significantly associated with a longer duration of exclusive breastfeeding. This is difficult to explain except as an artifact produced by the small sample size, but it was interesting that when we examined the birth month of
the children we found that 5 (41.7%) of 12 children with single infections and 2 (25%) of 8 children with multiple infections were born between April and July, the hottest months of the year, when mothers tend give other fluids in addition to breast milk. The finding of a difference in duration of exclusive breastfeeding did not appear to be related to nutritional status because there were no significant differences in WAZ or HAZ scores between the two groups at the time of weaning. A more detailed study will be required to assess nutritional and environmental factors and identify whether the multiple episodes of cryptosporidiosis result in growth deficits, or whether more episodes of cryptosporidiosis occurred in malnourished or more highly exposed children.

There were a number of limitations of this study, the greatest being the small sample size. Another limitation is that although we examined samples collected over a two-year period, the samples were only from children identified by microscopy to have previous symptomatic infections. There were also some occasions when the study child was unavailable for sample collection. Thus, the duration of every episode of cryptosporidiosis could not be estimated. Finally, although the infecting species could be identified in most episodes of cryptosporidiosis, it was not possible to identify the subgenotype in most asymptomatic infections, possibly because of a low parasite load.

Our current efforts are focused on a longitudinal investigation of cryptosporidiosis in a large birth cohort of children followed from birth to three years of age by using molecular techniques to detect, characterize, and quantify single and multiple asymptomatic and symptomatic Cryptosporidium spp. infections in this area. These data can then be correlated with clinical, nutritional, genetic, environmental, and immunologic parameters to determine risk factors for acquiring asymptomatic and symptomatic infections and re-infections with Cryptosporidium spp.

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REFERENCES


