Geographic Variation in the Sensitivity of Recombinant Antigen-based Rapid Tests for Chronic Trypanosoma cruzi Infection


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Abstract. Chagas disease affects 8–11 million people throughout the Americas. Early detection is crucial for timely treatment and to prevent non-vectorial transmission. Recombinant antigen-based rapid tests had high sensitivity and specificity in laboratory evaluations, but no Peruvian specimens were included in previous studies. We evaluated Stat-Pak and Trypanosoma Detect rapid tests in specimens from Bolivia and Peru. Specimens positive by three conventional assays were confirmed positives; specimens negative by two or more assays were confirmed negatives. In Bolivian specimens, Stat-Pak and Trypanosoma Detect tests were 87.5% and 90.7% sensitive, respectively; both showed 100% specificity. Sensitivity in Peruvian specimens was much lower: 26.6–33.0% (Stat-Pak) and 54.3–55.2% (Trypanosoma Detect); both had specificities > 98%. Even in Bolivian specimens, these sensitivities are inadequate for stand-alone screening. The low sensitivity in Peru may be related to parasite strain differences. Chagas disease rapid tests should be field tested in each geographic site before widespread implementation for screening.

INTRODUCTION

Chagas disease, caused by the parasite Trypanosoma cruzi, affects an estimated 8–11 million people throughout the Americas, and > 100 million people live in areas where they are at risk of becoming infected.1-3 Of those infected, as many as 30% develop debilitating—and potentially life-threatening—cardiac and/or gastrointestinal disease.4 People become infected when the T. cruzi-containing feces of the blood-sucking triatomine vector contaminate the bite wound or mucous membranes. Non-vectorial transmission may occur through blood transfusion, organ transplantation, laboratory accident, or congenitally. Without successful treatment, infection is life long. Therapy with currently available antitrypanosomal drugs is associated with reversion to negative serology in 50–60% of treated children5,6 and may decrease the likelihood of clinical progression of Chagas cardiomyopathy in adults.7 Early detection is crucial for timely treatment and to prevent non-vectorial transmission.

Conventional serologic tests, including the indirect hemagglutination assay (IHA), enzyme-linked immunosorbent assay (ELISA), and immunofluorescent antibody test (IFA), are commonly used to diagnose chronic Chagas disease. Because no single test has optimal sensitivity and specificity, the World Health Organization recommends the use in parallel of at least two distinct serologic assays to confirm T. cruzi infection.1 Epimastigote lysate-based ELISAs tend to have the highest sensitivity but have substantial rates of cross-reaction with other infections such as leishmaniasis.8,9 Furthermore, conventional assays require specialized laboratory equipment and trained laboratory personnel, and testing may require several days. Rapid tests based on recombinant antigens have been developed in an effort to improve the specificity of Chagas testing and to offer a simple diagnostic technique that provides rapid results. One commercially available rapid test based on several recombinant antigens, the Stat-Pak (Chembio Diagnostic Systems, Medford, NY), showed high sensitivity and specificity in laboratory testing of specimens from several endemic countries, including Bolivia.10,11 However, no specimens from Peru were included in the studies published to date. At the time we undertook this study, no studies on the performance of other rapid tests and no field evaluation of Stat-Pak had been published. To assess the utility of these tests for use in field studies, we evaluated the performance of two Chagas disease rapid tests in serum specimens from Peru and Bolivia.

MATERIALS AND METHODS

The specimens included in this analysis were collected during two epidemiologic studies of Chagas disease. The Bolivian specimens were tested in the study hospital laboratory by a technician used by the research project following the manufacturer’s instructions. Because the performance of the rapid tests in Peru was entirely unknown, the Peru specimen evaluation followed a stringently timed protocol, including readings by two independent observers, and was conducted in the Parasitic Diseases Serology Laboratory at the Centers for Disease Control and Prevention (CDC), Atlanta, Georgia. Both study protocols were approved by the Institutional Review Boards of the CDC, Johns Hopkins University Bloomberg School of Public Health, and Asociación Benéfica Proyectos en Informática, Salud, Medicina y Agricultura (PRISMA), Lima, Peru. Additional ethical approvals were obtained from the Hospital Universitario Japonés, Santa Cruz, Bolivia, for the study in Bolivia and the Instituto Nacional de Salud, Lima, Peru, for the Peru study.

Source and evaluation of Bolivian specimens. The Bolivian specimens were collected during recruitment for a study of congenital Chagas disease transmission in the Hospital Universitario Japonés, Santa Cruz, Bolivia, conducted from November 2006 to June 2007. Women presenting in labor to the obstetrical service of the hospital were asked to participate in the first step of the study, the aim of which was to...
identify women with *T. cruzi* infection at risk of transmission to their infants. After informed consent, a 5-mL blood specimen was collected from each woman, centrifuged in the hospital laboratory, and separated into clot and several aliquots of serum. The study laboratory technician used one fresh serum aliquot to perform the Stat-Pak and Trypanosoma Detect (InBios International, Seattle, WA) rapid tests in the hospital laboratory within a few hours of collection, following the manufacturer’s instructions. The rapid tests were used before their expiration dates. The Stat-Pak tests were donated by the Bolivian National Chagas Disease Control program and were used with consecutive specimens until the supply ran out; therefore, Stat-Pak testing was available for a subset of specimens only. The Trypanosoma Detect, which was not commercially available, was a prototype (ITC-8.2) provided by the manufacturer.

One aliquot of each serum specimen was frozen without additives and shipped from Bolivia to the Infectious Disease Laboratory at the Universidad Peruana Cayetano Heredia in Lima, Peru, where an in-house IFA following standard methods and two commercially available ELISAs (Chagatek ELISA; bioMerieux, Marcy l’Etoile, France [based on epimastigote lysate antigen] and Chagatest Recombinate ELISA [recombinant proteins]; Wiener Laboratories, Rosario, Argentina) were performed on all specimens. Following the manufacturer’s instructions for the Chagatek plates, the cut-off was set at 0.100 optical density units (OD) above the mean absorbance of the two negative control specimens included on each plate. For the Wiener plates, the cut-off was set at 0.300 OD units above the mean absorbance of three negative control specimens; specimens with OD values within the range of the cut-off ±10% were considered inconclusive and were re-run. If the OD still fell in this range, the final result was considered inconclusive.

Specimens positive or negative by all three conventional serologic tests were considered to have confirmed results. Specimens positive or negative by all three conventional serologic tests were considered inconclusive and were re-run. If the OD still fell in this range, the final result was considered inconclusive. Specimens positive or negative by all three conventional serologic tests were considered to have confirmed results. Specimens with discordant or inconclusive results were excluded.

**Source and evaluation of Peruvian specimens.** The Peruvian field site comprised two districts on the outskirts of the city of Arequipa where recent vector-borne *T. cruzi* transmission had been documented. Household-level insecticide spray campaigns were carried out by the Ministry of Health in late 2004; a serosurvey for Chagas disease was conducted in November–December 2005. Most participants were recruited in the public schools serving the two districts. In addition, the entire population of one hillside shantytown was invited to enroll in a community-based study. School age children living in the shantytown were recruited either in schools or in the community. A 5-mL blood specimen (3 mL for children < 5 years of age) was collected from each participant, separated into serum and cells, aliquoted into cryovials, and frozen without additives in the laboratory of the Universidad Nacional de San Agustín in Arequipa. Specimens were shipped on dry ice to Lima, Peru, for further processing.

All Peruvian specimens were tested by the same *T. cruzi* epimastigote lysate-based ELISA kit (Chagatek), as for Bolivian samples. Frozen aliquots of all ELISA-positive specimens plus a 10% random sample of ELISA-negative specimens were shipped on dry ice to CDC in Atlanta, and tested by an in-house IFA using standard methods; a titer of 1:32 was considered the threshold for a positive result. All specimens positive by ELISA or IFA, plus 20% of the negative specimens tested at CDC, were frozen and shipped on dry ice to the laboratory of one of the authors (LVK), where they were tested by the radioimmune precipitation assay (RIPA). The RIPA is often used in the United States to confirm positive results obtained in other tests and to resolve the status of specimens with discordant conventional tests. Specimens tested by all three assays (at least two assays for ELISA-negative specimens) were eligible for inclusion in this analysis. Specimens positive by all three assays (ELISA, IFA, and RIPA) were considered confirmed positive. Not all specimens negative by ELISA and IFA were tested by RIPA, because of the high cost of this assay; specimens were considered confirmed negative if they had negative results in the two (ELISA and IFA) or three assays (ELISA, IFA, and RIPA) used.

Confirmed positive and negative specimens were tested by the Stat-Pak and Trypanosoma Detect rapid tests in the CDC laboratory. For both tests, the manufacturer’s instructions were rigorously followed. Each test was read at 15 minutes based on a laboratory timer by two independent observers blinded to each other’s reading. The presence of a weak or strong line in the reaction zone was recorded as positive; its absence was recorded as negative. Readers also recorded comments, such as “weak positive,” “very weak positive,” “very faint,” or “unreadable.” For sensitivity/specificity analysis, interpretations of “positive,” “weak positive,” “very weak positive,” and “very faint” were all considered positive. Specimens with “unreadable” rapid test results excluded from analysis.

**Data analysis.** Sensitivities and specificities with 95% confidence intervals (CIs) and weighted κ coefficients for interobserver agreement were calculated using SAS V9.1 and SAS Enterprise Guide V4.1 (SAS Institute, Cary, NC). The Wilcoxon rank-sum test was used to compare differences for Chagatek ELISA absorbance (as measured by OD values minus cut-off value) for positive specimens, stratified by country of origin and rapid test results (true positive versus false negative).

**RESULTS**

Characteristics of the study populations, specimens, and testing are summarized in Table 1. In both cases, the case definition for confirmed positive specimens required positive results on three conventional serologic tests.

**Bolivian field assessment.** Of 530 specimens from women presenting to give birth in the hospital in Santa Cruz, 151 (28.4%) were confirmed positive and 365 (68.9%) were confirmed negative by all three conventional tests (Chagatek ELISA, Wiener ELISA, and IFA). Fourteen (2.6%) specimens with discordant (nine) or inconclusive (five) serologic results were excluded from the analysis. The Trypanosoma Detect was positive in 137 of the 151 confirmed positive sera and negative in all 365 confirmed negative sera, corresponding to a sensitivity of 90.7% and specificity of 100% (Table 2). The Stat-Pak was positive in 21 of 24 confirmed positive sera and negative in all 69 confirmed negative sera tested, yielding a sensitivity of 87.5% and specificity of 100%.

Six Stat-Pak readings carried the comment “weak positive” and one “very weak positive,” representing 30% of all positive readings; all were considered positive following the manufacturer’s instructions, and all were confirmed positive in the conventional tests. Two specimens had Stat-Pak–negative readings at 15 minutes but “very weak positive” at 25–30 minutes; based on the manufacturer’s instructions, these readings
were considered negative. One of these specimens was confirmed positive and the other negative based on the three conventional tests. Fifteen Trypanosoma Detect readings, or 10% of positive readings, were recorded as "weak positive," three of which were also "weak positive" on Stat-Pak. All were positive on the three conventional serologic tests.

**Evaluation in Peruvian specimens.** The serosurvey in Peru included 2,254 specimens, of which 120 (5.3%) were positive by the screening ELISA. Of these, 364 (120 positive and 244 negative by the screening ELISA) were shipped to CDC for confirmatory testing. At CDC, 102 (85.0%) of 120 ELISA-positive specimens and 5 (2.0%) of 244 ELISA-negative specimens were positive by IFA. Four ELISA-positive specimens lacked sufficient remaining volume to perform RIPA. RIPA results were therefore available for 98 of the 102 specimens positive by ELISA and IFA; 97 (99.0%) were positive, whereas 1 (1.0%) had an indeterminate RIPA result and was excluded from further analysis. Of 240 specimens negative by ELISA and IFA, 48 were tested by RIPA; 46 (95.8%) were negative. One (2.1%) was positive by RIPA, and one (2.1%) had an indeterminate result; these specimens also were excluded from further analysis. The group of specimens used to evaluate rapid test performance therefore consisted of 97 specimens positive by ELISA, IFA, and RIPA and 238 specimens negative by ELISA and IFA, 46 of which also were confirmed negative by RIPA (Table 1).

The observers reading the rapid tests in Peruvian specimens reported many weak and indeterminate readings for both tests. Although the tests are intended to be read as "positive" or "negative," 100 (59%) of the 169 positive test readings from two readers carried notations of "weak positive," "very weak positive," or "very faint" on one or both rapid tests (Figure 1). Weighted \( \kappa \) coefficient for interobserver agreement was 0.760 (95% CI: 0.652–0.867) for Stat-Pak, reflecting "substantial" agreement between readers and 0.833 (95% CI: 0.780–0.885) for Trypanosoma Detect, or "almost perfect" agreement between readers. The Stat-Pak showed sensitivities of 26.6% by Observer 1 and 33.0% by Observer 2; the specificity by both readers was 99.6% (Table 3). Trypanosoma Detect sensitivities were 53.8% and 54.6%; specificities were 98.7% and 98.3%.

### Table 1

<table>
<thead>
<tr>
<th>Source</th>
<th>Bolivia</th>
<th>Peru</th>
</tr>
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<tbody>
<tr>
<td>Population sampled</td>
<td>Pregnant women</td>
<td>All ages, school-aged children over-represented</td>
</tr>
<tr>
<td>Mean age in years (range)</td>
<td>24.7 (13–45)</td>
<td>16.2 (2–68)</td>
</tr>
<tr>
<td>Female sex</td>
<td>100%</td>
<td>52%</td>
</tr>
<tr>
<td>Specimen collected</td>
<td>5 mL blood</td>
<td>5 mL blood*</td>
</tr>
<tr>
<td>Rapid tests and location</td>
<td>Chagastek epimastigote lysate ELISA (UPCH§)</td>
<td>Chagastek epimastigote lysate ELISA (UPCH§)</td>
</tr>
<tr>
<td></td>
<td>IFA (UPCH§)</td>
<td>IFA (CDC‡)</td>
</tr>
<tr>
<td>Case definitions</td>
<td>Confirmed positive specimen: Positive on all three tests performed</td>
<td>Confirmed positive specimen: Positive on all three tests performed</td>
</tr>
<tr>
<td></td>
<td>Confirmed negative specimen: Negative on all three tests performed</td>
<td>Confirmed negative specimen: Negative on all tests performed (two or three)**</td>
</tr>
<tr>
<td>Eligible specimens††</td>
<td>530</td>
<td>358</td>
</tr>
<tr>
<td>Confirmed positive</td>
<td>191</td>
<td>97</td>
</tr>
<tr>
<td>Confirmed negative</td>
<td>339</td>
<td>236</td>
</tr>
</tbody>
</table>

* Three milliliters of blood for children < 5 years old.
† Hospital Universitario Japonés, Santa Cruz, Bolivia.
‡ Centers for Disease Control and Prevention, Atlanta, Georgia.
§ Universidad Peruana Cayetano Heredia, Lima, Peru.
¶ University of Iowa, Iowa City, IA.
** If RIPA was run; specimens with negative results on Chagastek ELISA and IFA were considered confirmed negative, if RIPA was not performed.
†† Eligible specimens were those tested by three conventional serologic assays (at least two assays for negative specimens from the Peru study). Fourteen Bolivian specimens were excluded (nine with discordant and six with inconclusive results). Twenty-three Peruvian specimens were excluded (20 with discordant and 3 with inconclusive results).

### Table 2

<table>
<thead>
<tr>
<th>Conventional serology</th>
<th>Chembio Stat-Pak</th>
<th>InBios Trypanosoma Detect</th>
</tr>
</thead>
<tbody>
<tr>
<td>Negative</td>
<td>69</td>
<td>365</td>
</tr>
<tr>
<td>Positive</td>
<td>3</td>
<td>14</td>
</tr>
</tbody>
</table>

| Sensitivity† | 87.5 (67.6–97.3) | 90.7 (84.9–94.8) |
| Specificity† | 100 (94.8–100) | 100 (99.0–100) |

* A lysate-based ELISA, a recombinant ELISA, and radioimmune precipitation assay were used to determine the final status of the specimens.
† Sensitivity or specificity expressed as a percentage and 95% CI.

**Figure 1.** The InBios Trypanosoma Detect (left) and Chembio Chagas Stat-Pak (right) rapid tests. Note the weak positive results in each of the tests. This figure appears in color at www.ajtmh.org.
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ELISA results. The ELISA absorbance among confirmed positive specimens was significantly higher in Bolivian than Peruvian specimens (0.842 versus 0.305; \( P < 0.0001 \); Figure 2). For Trypanosoma Detect, Bolivian specimens with true-positive results had a median absorbance of 0.873 versus 0.203 for false negatives (\( P < 0.0001 \)); in Peruvian specimens, the respective values were 0.394 and 0.195 (\( P < 0.0001 \); Figure 3). For Stat-Pak, the median for Bolivian specimens with true-positive results was 0.831 compared with 0.122 for false negatives (\( P = 0.08 \)); in Peruvian specimens, the respective values were 0.470 and 0.254 (\( P < 0.05 \); Figure 4).

DISCUSSION

Our evaluation of the Chembio Stat-Pak and InBios Trypanosoma Detect rapid tests in specimens from Bolivia and Peru showed highly variable sensitivities. In Peruvian specimens, both rapid tests had sensitivities < 60%, making them unusable in this setting in their current forms. Although the sensitivities in Bolivian specimens were much higher than in Peruvian samples, they still fell short of previously published values, 10,11,18 missing 10% or more of specimens confirmed positive by conventional serology. In both locations, the ELISA absorbance values were significantly higher among specimens with true-positive rapid test results than among those with false-negative results, suggesting that the rapid test detection thresholds may be set too high. In Peru, many rapid test readings were noted as “very weak” and therefore difficult to interpret. Nevertheless, interobserver agreement was high. The specificities determined in specimens from both locations were acceptably high.

The poor sensitivity of the Stat-Pak test, particularly in the Arequipa specimens, contrasts sharply with previously published data on its performance. One evaluation of Stat-Pak showed sensitivities of 98.5% in 393 Brazilian specimens and 100% in 352 specimens from Honduras, Argentina, Venezuela, and Bolivia. 10 An evaluation of the Stat-Pak in nearly 6,000 blood bank specimens from Honduras, El Salvador, and Nicaragua showed sensitivity of 99.6%. 11 The Stat-Pak was found to have a somewhat lower sensitivity of 93.4% in a recent field study in Sucre, Bolivia. 18 The authors hypothesized that the lower sensitivity resulted from the use of whole blood instead of serum; however, no results using serum from the same study subjects were provided to address this issue. Another recently published study from southern Bolivia reported a 5.9% discordance rate between Stat-Pak and an epimastigote lysate-based ELISA (Wiener Chagatest), but no details were provided regarding the rate of false-positive and false-negative results. 19

Our findings suggest that there may be more regional variation in antibody response to T. cruzi infection than is usually acknowledged. 20 In a study using ELISAs based on each of

![Figure 2](image2.png)

**Figure 2.** Distribution and median of Chagatek ELISA absorbance (OD minus cut-off value) among confirmed positive samples in studies in Bolivia and Peru. Confirmed positive specimens were defined as those with positive results on all three conventional serologic tests (Chagatek ELISA, IFA, and either Wiener Recombinant ELISA [Bolivia] or RIPA [Peru]).

![Figure 3](image3.png)

**Figure 3.** Distribution and median of Chagatek ELISA absorbance (OD minus cut-off value) of confirmed positive samples with true-positive and false-negative results by Trypanosoma Detect rapid test in studies in Bolivia and Peru.
We were unable to use the same second confirmatory test (RIPA) in Bolivian as Peruvian specimens because of resource constraints. Nevertheless, we believe that our definition of confirmed positive was equally stringent for both sets of specimens. Indeed, the use of three tests may have excluded positive specimens with lower antibody levels and thereby resulted in an overestimate of the true sensitivity of the rapid tests. The only methodologic difference that might contribute to a lower sensitivity in the Peruvian specimens was the fact that they had been through two freeze-thaw cycles before the rapid tests were used, whereas the Bolivian specimens were tested fresh. However, RIPA testing of the Peruvian specimens occurred after a third freeze-thaw cycle and showed high concordance with the screening ELISA. In any case, two freeze-thaw cycles should not cause a 50–70% decrease in the sensitivity of a robust serologic test. Clearly, further evaluations of rapid test performance in a broader range of populations and under a variety of conditions are still needed.

Rapid tests for Chagas disease have the potential to facilitate both prenatal screening and surveys to detect infected children who would benefit from drug treatment. To date, the challenging logistics of community and hospital-based screening and follow-up have prevented these efforts from reaching large proportions of the population in need. The Stat-Pak and Trypanosoma Detect rapid tests are simple to use and require no refrigeration, specialized equipment, or highly trained laboratory personnel. The results are available within 15 minutes, eliminating the need to return a second time to a community or to track down an infected woman after discharge from the hospital. The use of recombinant antigens rather than whole epimastigote preparations may improve the specificity of Chagas testing, particularly in areas where Chagas disease and leishmaniasis are co-endemic. These attributes make rapid tests very appealing for use in field epidemiology and clinical diagnostics. Stat-Pak has been used for community-based screening of children in Bolivia, and its use has been advocated for other settings where conventional serology is not readily available.

However, the specific antigens and format used in rapid tests may result in varying sensitivity depending on the location in which the test is applied: Arequipa seems to be one such location for which these rapid tests are not well adapted. Furthermore, rapid test sensitivity in Bolivian specimens was low for use as the sole screening tool. A child or pregnant woman with a false-negative rapid test may not ever be tested again, but a false-positive rapid test result will be detected when confirmatory serology is run. Thus, higher sensitivity would be worth the trade-off of lower specificity that would inevitably result from lowering the detection threshold. Until consistently high sensitivity can be shown, rapid diagnostic tests should be carefully evaluated in specimens from the intended target locations before widespread implementation, and screening of new populations should continue to include testing by sensitive conventional assays in all specimens.

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