COMPARISON OF ANTIGEN-CAPTURE ELISA TO STOOL-CULTURE METHODS FOR THE DETECTION OF ASYMPTOMATIC ENTAMOEBA SPECIES INFECTION IN KAFER DAOUĐ, EGYPT

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Abstract. We performed a prospective field study in the village of Kafer Daoud in Menoufia, Egypt to compare the fecal culture method with enzyme linked immuno assay (ELISA) for detection of 170 kDa lectin antigen in feces for diagnosis of asymptomatic Entamoeba histolytica and Entamoeba dispar infection. All subjects with E. histolytica or E. dispar infection detected by culture also had positive ELISA for amebic antigen in their feces and an additional 5% Entamoeba infections missed by culture were detected by ELISA (P < 0.001 compared to culture). The presence of fecal anti-lectin IgA antibodies and serum anti-LC3 (recombinant cysteine-rich lectin protein) IgG antibodies were positive predictors for E. histolytica infection (P < 0.03). Of interest, infection with Trichomonas hominis but not Blastocystis hominis was positively associated with E. histolytica infection (P < 0.05). In conclusion, ELISA for detection of fecal lectin antigen is a more sensitive method than fecal culture for detecting asymptomatic E. histolytica infection.

INTRODUCTION

Entamoeba species, histolytica and dispar, infect up to 10% of the world’s population. The highest burden of Entamoeba sp. infection occurs in developing countries, with a combined prevalence as high as 25% in Mexico, and 38% in the continent of India, and 25% in sub-Saharan and tropical Africa. All E. dispar infections and 90% of E. histolytica intestinal infections are asymptomatic; E. histolytica causes approximately 50 million cases of invasive amebiasis (colitis and liver abscess) and 100,000 deaths per year. Based upon studies utilizing culture of fecal samples for detection, the prevalence of Entamoeba sp. infection in an endemic area was estimated at 2.6%. Culture provides up to a four-fold increase in sensitivity compared to microscopy for detection of E. histolytica trophozoites or cysts in feces; however, its absolute sensitivity is unknown. Utilizing polyclonal or uncharacterized monoclonal anti-amebic antibodies, ELISA has been successful in detecting E. histolytica antigen in stool. However, early reports did not differentiate E. histolytica from E. dispar infection. Monoclonal antibodies to the galactose-inhibitable adherence lectin 170 kDa subunit have been utilized in ELISA to differentiate E. histolytica from E. dispar. The epidemiologic studies using Tech lab kits demonstrated lower sensitivity than the gold standard of culture and isoenzyme analysis. Detection of serum lectin antigen by ELISA has been demonstrated to be useful in identifying E. histolytica infections. In a prospective field study in a small village in Egypt, we investigated whether stool culture or a monoclonal antibody-based ELISA for detection of fecal-lectin antigen method was more sensitive for diagnosing asymptomatic E. histolytica and E. dispar infection.

MATERIALS AND METHODS

Study populations. Single serum and stool samples were obtained over a one-month period from each of the 182 randomly selected subjects, two-thirds of whom were women living in Kafer Daoud village located in Menoufia, Egypt. Subjects ranged in age from 6 to 60 years and were asymptomatic, without recent history of gastrointestinal complaints or use of amebicidal drugs such as metronidazole. Fecal samples were also collected from 55 patients at the University of Minnesota Hospital, Minneapolis, Minnesota, with no history of parasitic infection and negative fecal microscopy; control sera were obtained previously from 47 healthy employees at the University of Virginia, Charlottesville, Virginia, with no history of amebic infection. Entamoeba species, Blastocystis hominis and Trichomonas hominis were differentiated by microscopy.

Stool culture. Fecal samples were collected in the morning and kept in ice until transported to Cairo within the next twelve hours. Samples were cultured in Robinson’s Medium as described. After 48 hours, an aspirate of the starch layer was mixed with double strength Lugol’s iodine and examined microscopically. A second reading was performed after 96 hours. Positive cultures were subcultured every 2 or 3 days using fresh slope with starch, 2 drops of peptone, 2 drops of erythromycin and BRS diluted 1 in 4 with pthalate filled up to the bottle neck for incubation at 37°C.

Hexokinase isoenzyme electrophoresis. Entamoeba species were differentiated by electrophoretic migration of hexokinase isoenzymes as described by Sargeant.

ELISA for detection of 170 kDa lectin antigen in serum and feces. ELISA for detection of 170 kDa antigen was performed according to the method of Abd-Alla and others. Briefly, 96 well flat-bottomed polystyren microtiter ELISA plates (Coster, Corning, NY) were coated with monoclonal antibody 3F4, which recognizes lectin epitopes present in both E. histolytica and E. dispar, or with 8C12 antibodies, which recognizes epitopes found only in E. histolytica lectin. Non-reactive sites in the wells were blocked with 1% BSA in coating buffer. Stool samples were mixed in an equal volume of PBS containing 2 mM PMSF (United States Biochemical Corp., Cleveland, OH), serum samples were diluted 1:100 in PBS-Tween 1% BSA. Serum or stool samples were added at 100 μl/well and incubated for 2 hours at room temperature or overnight at 4°C. Alkaline phosphatase conjugated anti-lectin monoclonal antibodies (8A3, which recognizes both E. histolytica and E. dispar, or 1G7, specific for E. histolytica) were added at 1:1000 dilution. Developing, reading (Bio-Rad Microplate reader-Benchmark, Hercules, CA) and correction for nonspecific background were performed as described.
ELISA for detection of fecal anti-lectin IgA antibodies. 
ELISA for the detection of fecal anti-lectin IgA antibodies was performed as described by Abo-El-Maged and others. Briefly, ELISA plates were coated with a purified recombinant cysteine-rich portion of the lectin’s 170 kDa subunit (LC3) (0.2 μg protein/well) and feces were processed as above except that alkaline phosphatase-conjugated goat anti-human IgA antibodies (Sigma, St, Louis, MO) were added (1:2000) in PBS-Tween 1% BSA. Developing, reading and correction of nonspecific background-binding were performed as described.

ELISA for detection of serum anti-LC3 IgG and IgA antibodies. ELISA for detection of serum anti-LC3 IgG and IgA antibodies were performed as described. Briefly, serum samples were diluted at 1:1,000 for assay of IgG and 1:500 for IgA determination in PBS-Tween containing 1% BSA. Alkaline phosphatase-conjugated goat anti-human IgG antibodies diluted in PBS-Tween with 1% BSA (1:5,000 for IgG and 1:2,000 for IgA) were utilized as secondary antibodies. Developing, reading and correction of nonspecific background-binding were performed as described.

Statistics. All results were expressed as percent positive and percent negative. Z test (converted to P value) was used to determine the significance of difference. The Z test is defined as: Z = (P1 - P2) / √ P(Q / N1) – (Q / N2) where P1 = proportion of positive in group one, P2 = proportion of positive in group two, P = pooled proportional estimate = (X1 + X2) / (N1 + N2), N1 = number of positive in group one, N2 = number of positive in group two, N1 = total number of group one, N2 = total number of group two, Q = (1 - P). In the current study the two samples Z test was used.

RESULTS

Stool culture and isoenzyme determination. Fifteen of 182 Egyptian subjects studied (8.2%) had a positive stool culture for *Entamoeba* species. Of these, by zymodeme determination eight were found to be *E. dispar*, five were *E. histolytica* and two were of indeterminate species. Twenty three (12.6%) of the 182 subjects had a positive microscopy for amebic cysts; other parasites found included Giardia lamblia (1), Entrobius vermiculari (7), Ascaris lumbercoid (2), Ancylostom duodena (2), Hymenolypus nana (1), Trichuris trichiura (1), and Schistosoma mansonian (3).

ELISA for detection of fecal 170 kDa lectin antigen. As summarized in Table 1, 100% of stool samples positive by culture for *E. histolytica* or *E. dispar* species were correctly identified and differentiated by ELISA used to detect fecal lectin antigen. In the two positive cultures whose species were not identifiable by zymodeme analysis, we found that up to 100 trophozoites per well from these cultures were not reactive in the ELISA. Of 23 fecal samples positive for microscopy, 84% were positive for fecal lectin antigen detection (41% *E. histolytica*, 43% *E. dispar*). Of the 17 infected with other parasites detected by microscopy, only 3 had positive lectin antigen ELISA tests. As reflected in Table 1, additional *E. histolytica* and *E. dispar* infections were detected by ELISA, (P < 0.001) compared to stool culture.

**Table 1**

<table>
<thead>
<tr>
<th>ELISA for detection of Entamoeba histolytica (E.h.) and Entamoeba dispar (E.d.) lectin antigen</th>
<th>E.h. antigen (+) (n = 39)</th>
<th>E.d. antigen (+) (n = 44)</th>
<th>Antigen (-) (n = 99)</th>
</tr>
</thead>
<tbody>
<tr>
<td>E.h. culture (+) (n = 5)</td>
<td>5 (100%)</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>E.d. culture (+) (n = 8)</td>
<td>0</td>
<td>8 (100%)</td>
<td>0</td>
</tr>
<tr>
<td>Fecal culture (-) (n = 167)</td>
<td>34 (34/167)</td>
<td>36 (36/167)</td>
<td>97 (97/167)</td>
</tr>
</tbody>
</table>

* = positive; – = negative.

This study compares the use of stool culture and zymodeme analysis to performance of ELISA for fecal lectin antigen to detect *E. histolytica* and *E. dispar* infections in a highly endemic area. Results were correlated with fecal microscopy, ELISA for detection of serum 170 kDa lectin.

**DISCUSSION**

This study compares the use of stool culture and zymodeme analysis to performance of ELISA for fecal lectin antigen to detect *E. histolytica* and *E. dispar* infections in a highly endemic area. Results were correlated with fecal microscopy, ELISA for detection of serum 170 kDa lectin.
antigen, serum and mucosal anti-amebic antibody responses, and by culture, with co-infection by B. hominis and T. hominis.

The prevalence of E. histolytica and E. dispar infection as determined by stool culture was in agreement with reports from another endemic area.10 It is unclear why culture has such low sensitivity, compared to ELISA for fecal lectin antigen detection. The high specificity of the monoclonal antibodies used in the antigen-capture ELISA was clearly demonstrated. All positive cultures for E. histolytica or E. dispar were correctly identified and differentiated by the ELISA. Moreover, the two positive cultures for non-histolytica and non-dispar trophozoites (hexokinase isoenzyme electrophoresis revealed three bands) were negative by ELISA.

Using ELISA to detect fecal lectin antigen we found that E. histolytica or E. dispers infection were almost ten-fold more prevalent than estimated by fecal culture. The overall prevalence of E. dispers infection might actually be higher than reported, as the monoclonal antibodies used are unable to detect mixed infections in subjects positive for E. histolytica fecal antigen. The current study confirmed that E. histolytica, but not E. dispers, infection induces serum anti-amebic antibody responses, as reported in many studies worldwide.11–14,22,23

The Kafer Daoud villagers are considered to be at high risk for gastrointestinal parasitic infections due to the absence of proper sanitary disposal, their continuous contact with domestic animals, their having pets who lack veterinary care, and a high density of flies year around. Blastocystis hominis is a common human parasite residing in the colon. Many reports support a role for B. hominis as a potential pathogen causing diarrhea, abdominal discomfort, anorexia, and flatulence.24,25 However, it is controversial whether B. hominis is a pathogen; this is especially of concern in immunocompromised individuals.26–28 The prevalence of B. hominis has been found to be as high as 61.8% in a riverside community,29 51.8% of children in Chile,30 54% of villagers in New Guinea,31 and 49% of residents in a suburban community in Venezuela.32 In contrast, B. hominis was found in only 11.5% of asymptomatic subjects in the United States33 and 0.5% of asymptomatic subjects in Japan.34 Risk factors for infection include travel to the tropics,35 infection among other family members,36 and contact with pets.36 The high prevalence of asymptomatic B. hominis infection (52.2%) among the villagers in this study is consistent with reports from various parts of the world.26,27,29,30

**Table 3**

<table>
<thead>
<tr>
<th>Blastocystis hominis and Trichomonas hominis infection in stool screened for Entamoeba histolytica (E.h.) and Entamoeba dispar (E.d.) 170 kDa antigen</th>
</tr>
</thead>
<tbody>
<tr>
<td>B. hominis (+)</td>
</tr>
<tr>
<td>(n = 95)</td>
</tr>
<tr>
<td>14 (14.7%)</td>
</tr>
<tr>
<td>T. hominis (+)</td>
</tr>
</tbody>
</table>

Experimentally, up to 100 T. hominis and B. hominis trophozoites per well were not identified by the monoclonal antibodies used in ELISA. E. dispers is more prevalent than E. histolytica in B. hominis infection (P = 0.011), while E. histolytica is more frequent than E. dispers in T. hominis (P = 0.018).

**Trichomonas hominis** is a nonpathogenic protozoan parasite that resides in the colon and small intestine; it is rarely associated with diarrhea.32 and has been found outside its normal habitat in the respiratory tract,38 in meningitis following GI fistula,39 and in esophageal intramural pseudodiverticulosis.40 The significance of co-infection between E. histolytica and T. hominis and between E. dispar and B. hominis is unknown, but could be related to shared risk factors for infection and microbial interference in the gut. Importantly, the monoclonal antibodies used to detect Entamoeba sp. did not crossreact in ELISA with T. hominis or B. hominis. In addition, anti-B. hominis monoclonal antibodies are known to be nonreactive with E. histolytica antigen.41

In conclusion, we found that ELISA for detection of fecal 170 kDa lectin antigen is a more sensitive method than a single stool culture for diagnosis of asymptomatic Entamoeba species infection, indicating that asymptomatic E. histolytica infection is more common than previously appreciated. The finding of co-infection with nonpathogenic intestinal protozoans may provide further clues to the epidemiology of Entamoeba species infection and microbial interactions in the intestinal lumen.

Acknowledgments: The author would like to thank Joan Portel and Shana Brooks for expert secretarial assistance.

Financial support: This work was supported by a grant from AVI-CENNE Program, Commission of European Communities, A/V/–CENNE Program, Commission of European Communities, AVI–CT93–0008 to Dr. Abd-Alla and grants PO1-A136359 and UI01-A135840 from the National Institutes of Health to Dr. Ravdin.

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