INTRODUCTION
Post transplant parasitic infections are a rarity and occur in around 2% of transplant recipients; the intestinal helminth *Strongyloides stercoralis* (*Ss*) is found in contaminated soil in hot and humid tropical and subtropical regions of Africa, South and East Asia, and South America. Infective larvae from contaminated soil enter the host venous system from the skin to end up in the lungs and are then ingested into the GI tract by the pharynx where they mature into adults. Approximately half of the immune competent hosts become carriers and the remainder usually present with mild abdominal and respiratory symptoms. Transplant recipients receive steroids, have altered cellular immunity, and are at increased risk for hyperinfection syndrome because of an accelerated auto-infective cycle, rapid multiplication, and systemic larval dissemination. This can result in serious complications including GI bleeding, bacteremia, meningitis, liver abscesses, pneumonia, and death. We report a *Strongyloides* hyper-infection syndrome in a Saudi kidney recipient with recurrent *Streptococcus bovis* bacterial meningitis and significant weight loss 2 months after deceased donor kidney transplantation.

CASE REPORT
A 27-year-old Saudi male with end stage renal disease secondary to focal segmental glomerulosclerosis on regular hemodialysis for 4 years, received a 4 HLA antigen mismatch standard criteria deceased donor kidney from an expatriate Indian donor. Induction comprised anti-thymocyte globulin and methylprednisolone with immediate graft function after 8 hours of cold ischemia. He was discharged home on tacrolimus, mycophenolate, and prednisolone with a serum creatinine of 102 mmol/L. He was febrile (38.5°C) with tachycardia and pallor with mild signs of meningeal irritation. He also gave a history of abdominal pain, vomiting, and diarrhea for the preceding month associated with weight loss. A CT brain scan was reported as a paralytic ileus/sub-acute intestinal obstruction, CSF showed a high neutrophil count, however like the blood sample, was culture negative (Table 1). The impression was a recurrence of ABM for which he again received intravenous antibiotics with a reduction in the immunosuppression. The next day, he developed a cough with sputum, a chest examination and chest x-ray were unremarkable, but *Escherichia coli*, sensitive to cephalosporins was isolated from the sputum. In view of the weight loss and diarrhea, thyroid function tests were also requested, which revealed a slightly elevated T4 and a low thyrotropin (TSH). Thyroiditis was also suggested by the low uptake on the nuclear scan with a negative autoimmune screen, and he was started on carbimazole. The pp65 antigen and *Brucella* were negative and all tumor markers were also within normal. Mild eosinophilia in the differential count with a 10-day twice daily course of 400 mg albendazole treatment with a 10-day twice daily course of 400 mg albendazole with improvement in abdominal pain, vomiting, and diarrhea. The stool analyses had repeatedly been negative for *Ss* and no *O*+*P* were ever documented and the preoperative (before immunosuppression) eosinophil count was also found to be normal. The regular immunosuppression was resumed after completing the intravenous antibiotics. He regained 8 kg in the next 3 weeks and was back to his normal weight in 3 months. At 36 months, he remains symptom free with excellent renal function.

DISCUSSION
The intestinal parasite *S. stercoralis* is not endemic in Saudi Arabia; however, the majority of deceased donors are...
expatriates from endemic areas of South India, Sri Lanka, Indonesia, and the Philippines. Donor-linked parasitic transmission has been suspected in transplant recipients who received organs from donors from endemic areas or proven when different recipients developed Ss after receiving different organs from the same donor. In our case, it is unlikely that this infection was donor derived because the recipient of the sister kidney did not develop any symptoms, and neither a donor eosinophil count nor a donor serum sample was available to confirm Ss IgG antibodies with enzyme-linked immunosorbent assay (ELISA). Cyclosporine A has been reported to confer immunity against Ss but both kidney recipients in this case were receiving tacrolimus.

Expatriate workers in Saudi Arabia who become deceased donors from known Ss-endemic areas should be considered a transmission risk and predonation screening for this parasite and other imported infections would be beneficial. For Ss, screening with eosinophil count and IgG antibodies is desirable, but the unreliable history and short period of evaluation remains an obstacle for productive screening. This deceased donor was from an endemic area but the required screening was not performed and the use of steroids in donor preconditioning enhances the potential for Ss dissemination and hyperinfection. In addition, this donor received regular dexamethasone following the head injury, and the recipient was given bolus steroids as part of induction and both would have played a part in reactivating Ss, causing hyperinfection and dissemination in the recipient.

The source of this transmission remains a mystery, there being no direct evidence that this Ss infection was donor derived, first because our Saudi recipient had never traveled to an endemic area and second, the other Saudi who received the sister kidney also did not develop any symptoms. The majority of Ss cases present within 1–3 months after transplantation, and our recipient developed symptoms 2 months post transplant. Meningeal symptoms, the positive CSF culture and the E. coli from the sputum represent the bacteremia associated with unrestricted larval migration through the bowel wall that can result in a high incidence of S. bovis meningitis, especially after steroid therapy. Because mortality in transplant recipients who develop the hyperinfection syndrome is very high reaching up to 85%, it has been suggested that diagnosis in the donor should preclude transplantation. Multisystem involvement is not rare, as in our case, with paralytic ileus, E. coli acute tracheobronchitis, meningitis, and weight loss. Saudi Arabia is not considered endemic for Ss and may be the reason it was not considered in the differential diagnosis. Early diagnosis is vital but is liable to get delayed in non-endemic areas because the disease is neither suspected, nor pursued if the stool samples are negative, as in our case. Peripheral eosinophil counts may be low because of steroids and thus unhelpful in affected transplant recipients and diarrhea is generally considered to be drug related; our patient received high dose dexamethasone that might have masked the oesinophilia in the first admission. In case the diarrhea continues, it is important to test several fresh stool samples and an agar culture for better yields of the

<table>
<thead>
<tr>
<th>Test</th>
<th>First admission</th>
<th>Second admission</th>
</tr>
</thead>
<tbody>
<tr>
<td>CSF glucose</td>
<td>2.9</td>
<td>3.0</td>
</tr>
<tr>
<td>Blood glucose</td>
<td>6.3</td>
<td>6.5</td>
</tr>
<tr>
<td>Protein</td>
<td>0.5</td>
<td>1.0</td>
</tr>
<tr>
<td>White cells</td>
<td>34/cu mm</td>
<td>320/cu mm</td>
</tr>
<tr>
<td>Polymorphs</td>
<td>96%</td>
<td>97%</td>
</tr>
<tr>
<td>Lymphocytes</td>
<td>4%</td>
<td>3%</td>
</tr>
<tr>
<td>Cryptococcus antigen</td>
<td>Negative</td>
<td>Negative</td>
</tr>
</tbody>
</table>

*Leukocytes, overwhelmingly neutrophils, are present in both samples.*
parasite.\textsuperscript{13} Antibodies to \textit{Ss} using ELISA can confirm diagnosis, especially when repeated stool testing is negative.\textsuperscript{3} Weight loss is common in GI \textit{Ss} infection because of malabsorption,\textsuperscript{7,13} which in our recipient was ascribed to mild thyrotoxicosis, diarrhea, cytomegalovirus, and other prevailing infective phenomena. Because ivermectin, considered the drug of choice was not available,\textsuperscript{7,15,18} the recipient was given albendazole 400 mg for 10 days. As reactivation and relapse remains a possibility in immunosuppressed patients, some researchers have advised intermittent monthly therapy.\textsuperscript{19}

\textit{Strongyloides} hyperinfection was neither suspected nor confirmed in the first admission and the symptoms that led to urgent admission were primarily related to the central nervous system (CNS) along with mild abdominal symptoms with weight loss. The meningeal symptoms were overwhelming and overshadowed the abdominal symptoms, and the negative stool samples were further misleading. The focus remained on the CNS, especially when \textit{S. bovis} was isolated from the CSF confirming acute bacterial meningitis. It is thought that migration of larvae through the bowel wall resulted in the bacteremia\textsuperscript{6} that caused the first episode of \textit{Ss} infection. In retrospect, our recipient had abdominal symptoms and would have benefitted from an upper GI endoscopy, because \textit{Ss} were not diagnosed until its eventual discovery on duodenoscopy. The abdominal symptoms were present at the time of the first admission and had the patient been more cooperative, we could perhaps have diagnosed \textit{Ss} earlier, with duodenoscopy. During the second admission, the patient also developed a cough with \textit{E. coli}-positive sputum indicating, in retrospect perhaps, another wave of \textit{Ss} driven bacterial translocation. This is the main reason for the reported \textit{Klebsiella pneumonia} and \textit{S. bovis} acute bacterial meningitis, in patients with \textit{Ss}, especially those receiving steroids.\textsuperscript{11,12}

This however is the first report from Saudi Arabia of \textit{Strongyloides} hyperinfection syndrome with recurring ABM in a Saudi kidney recipient. Despite a relatively late diagnosis and the unavailability of ivermectin, this recipient had a good outcome.

In conclusion, \textit{Ss} is a serious disease and requires a high index of suspicion in non-endemic areas and all donors from endemic areas should be screened. Abdominal symptoms, eosinophilia, pruritis, or asthma in any transplant recipient should warrant examination of fresh stool samples along with ELISA testing for IgM and IgG antibodies. Treatment should be initiated when \textit{Ss} is suspected because delay in treatment can be fatal\textsuperscript{6,7,18} and should include ivermectin.\textsuperscript{6,7,18} Repeated stool examination and therapy are also recommended because relapses are common.\textsuperscript{19}

Received August 25, 2013. Accepted for publication December 2, 2013. Published online January 6, 2014.

Authors’ addresses: Taqi T. Khan, Riyadh Military Hospital, Renal Transplant Surgery, Riyadh, Saudi Arabia, E-mail: taqikhan@yahoo.com. Fatehi Elzein, Riyadh Military Hospital, Infectious Diseases, Riyadh, Saudi Arabia, E-mail: fatehielzein@yahoo.com. Abdullah Fiaar, Riyadh Military Hospital, Pathology, Riyadh, Saudi Arabia, E-mail: Faheem Akhtar. Faheem Akhtar, Riyadh Military Hospital, Transplant Nephrology, Riyadh, Saudi Arabia, E-mail: drfaheemakhtar@yahoo.com.

REFERENCES